

TRANSMITTAL LETTER TO THE UNITED STATES

ATTORNEY'S DOCKET NUMBER 0093/000003

DESIGNATED/ELECTED OFFICE (DOLEGO, S.,)
CONCERNING A FILING UNDER 35 U.S.C. 371

U.S. APPLICATION NO. (If known, see 37 CFR 1.5)

INTERNATIONAL APPLICATION NO. PEP 00/02701

ATENT & TRA

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INTERNATIONAL FILING DATE 12 OCT 2000

PRIORITY DATE CLAIMED

March 28, 2000

LE OF INVENTION: A NEW CLASS OF ENZYMES IN THE BIOSYNTHETIC PATHWAY FOR THE PRODUCTION OF TRIACYLGLYCEROL RECOMBINANT DNA MOLECULES ENCODING THESE ENZYMES

APPLICANT(S) FOR DO/EO/US Anders DAHLQUIST; Ulf STAHL; Marit LENMAN; Antoni BANAS Hans RONNE; Sten STYMNE

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

- 1. /X/ This is a FIRST submission of items concerning a filing under 35 U.S.C. 371.
- 2. / / This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371.
- 3. /X/ This express request to begin national examination procedures (35 U.S.C.371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
- A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
- 5. /X/ A copy of the International Application as filed (35 U.S.C. 371(c)(2)).
 - is transmitted herewith (required only if not transmitted by the International Bureau). has been transmitted by the International Bureau.

 - b.// is not required, as the application was filed in the United States Receiving Office (RO/USO).
- 6. /X/ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
- 7. / / Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)).

 - are transmitted herewith (required only if not transmitted by the International Bureau). have been transmitted by the International Bureau. have not been made; however, the time limit for making such amendments has NOT expired. b.//
 - have not been made and will not be made.
- 8: / / A translation of the amendments to the claims under PCT Article 19(35 U.S.C. 371(c)(3)).
- 9. /X/ An oath or declaration of the inventor(s)(35 U.S.C. 171(c)(4)).
- 10./ / A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).
- Items 11. to 16. below concern other document(s) or information included:
- 11./X/ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
- An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 12.// is included.
- 13./X/ A FIRST preliminary amendment.
 // A SECOND or SUBSEQUENT preliminary amendment.
- 14./ / A substitute specification.
- 15./ / A change of power of attorney and/or address letter.
- 16./X/ Other items or information.
 - International Search Report
 International Preliminary Examination Report



JC05 Rec'd PCT/PTO

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INTERNATIONAL APPLN. NO. PCT/EP 00/02701

ATTORNEY'S DOCKET NO. 0093/000003

17. /X/ The following fees are submitted	<u>chin</u>	CULATIONS	<u>FIO</u>	USE ONLY
BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5)):				
Search Report has been prepared by the				
EPO or JPO	\$860.00	860.00	1	
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International preliminary examination fee paid to	USPTO			
(37 CFR 1.482)	\$690.00		l	
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No international preliminary examination fee paid	l to			
USPTO (37 CFR 1.482) but international search fee				
to USPTO (37 CFR 1.445(a)(2))			ı	
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(37 CFR 1.482) nor international search fee	•			
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Surcharge of \$130.00 for furnishing the oath or o				
later than // 20 //30 months from the ear	llest			
claimed priority date (37 CFR 1.492(e)).				
<u>Claims</u> <u>Number Filed</u> <u>Number Extra</u>	Rate			
Total Claims 22 -20 2	X\$18.	36.00	ļ	
Indep.Claims 4 -3 1	х\$80.	80.00	i	
Multiple dependent claim(s)(if applicable)	+270.			
TOTAL OF ABOVE CALCULATION	=	116.00		
Reduction of & for filing by small entity, if app	licable.			
Verified Small Entity statement must also be file				
(Note 37 CFR 1.9, 1.27, 1.28).				
(NOCC 37 CIR 1.3) 1.27, 1.20).				
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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of)	
DAHLQUIST et al.)	BOX PCT
PCT/EP 00/02701 Intl. Filing Date: March 28, 2000)	,	
US Serial No.: TO BE ASSIGNED)	

For: A NEW CLASS OF ENZYMES IN THE BIOSYNTHETIC PATHWAY FOR THE PRODUCTION OF TRIACYLGLYCEROL AND RECOMBINANT DNA MOLECULES ENCODING THESE ENZYMES

Honorable Commissioner of Patents and Trademarks Washington, D.C. 20231

PRELIMINARY AMENDMENT

Sir:

Prior to examination of the above-identified U.S. National Stage application, kindly amend the application as follows.

CLEAN VERSION OF ALL CLAIMS

Cancel claims 1-27, all the claims in this case, and substitute new claims 28-49 as follows:

- 28. An enzyme, designated as phospholipid:diacylglycerol acyltransferase (PDAT), catalyzing in an acyl-CoA-independent reaction the transfer of fatty acids from phospholipids to diacylglycerol in the biosynthetic pathway for the production of triacylglycerol and comprising an amino acid sequence as set forth in SEQ ID NO:2 or a functional fragment, derivate, allele, homolog or isoenzyme thereof.
- 29. An enzyme according to claim 28, comprising an amino acid sequence encoded through a nucleotide sequence as set forth in SEQ ID NO:1 or a homologous nucleotide sequence which is at least about 40% identical to a nucleotide sequence of SEQ ID NO. 1.
- 30. An enzyme according to claim 28, comprising an amino acid sequence as set forth in SEQ ID NO. 16, 20 or 22, or a functional fragment, derivate, allele, homolog or isoenzyme thereof.
- 31. An enzyme according to claim 28, comprising an amino acid sequence selected from the group consisting of sequences as set forth in SEQ ID NO. 6, 8, 13, 14, 15, 17, 18, 25 and 27, a functional fragment, derivate, allele, homolog or isoenzyme thereof.
- 32. An enzyme according to claim 28, comprising an amino acid sequence encoded through a nucleotide sequence, a portion, derivate, allele or homolog thereof selected from the group consisting of sequences as set forth in SEQ ID NO. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 and 31, or a functional fragment, derivate, allele, homolog or isoenzyme of the enzyme encoding amino acid sequence.
 - 33. A nucleotide sequence according to claim 32, selected

from the group consisting of sequences as set forth in SEQ ID NO. 1, 3, 4, 10, 11, 19, 21, 23, 24, 29 and 30, or a portion, derivate, allele or homolog thereof.

- 34. A partial nucleotide sequence corresponding to a full length nucleotide sequence according to claim 32, selected from the group consisting of sequences as set forth in SEQ ID NO. 5, 7, 9, 12, 25, 26, 28 and 31, or a portion, derivate, allele or homolog thereof.
- 35. A nucleotide sequence according to claim 32, comprising a nucleotide sequence which is at least 40% identical to a nucleotide sequence selected from the group consisting of those sequences set forth in SEQ ID NO. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 and 31.
- 36. A gene construct comprising a nucleotide sequence as set forth in SEQ ID No. 1, or a homologous nucleotide sequence which is at least about 40% identical to the nucleotide sequence of SEQ ID No. 1, which is operably linked to a heterologous nucleic.
- 37. A vector comprising a gene construct according to claim 36, or the nucleotide sequence as set forth in SEQ ID No. 1, or a homologous nucleotide sequence which is at least about 40% identical to the nucleotide sequence of SEQ ID No. 1.
- 38. A vector according to claim 37, which is an expression vector.
- 39. A vector according to claim 37, further comprising a selectable marker gene and/or nucleotide sequences for the replication in a host cell or the integration into the genome of the host cell.
- 40. A transgenic cell or organism comprising one or more of the following:
- a) a nucleotide sequence a_1) to a_4),

- b) a gene construct b_1 , and
- c) a vector c_1),

wherein

- a_1) is a nucleotide sequence as set forth in SEQ ID NO. 1 or a homologous nucleotide sequence which is at least about 40% identical to a nucleotide sequence of SEQ ID NO. 1,
- a₂) is a nucleotide sequence, a portion, derivate, allele or homolog thereof selected from the group consisting of sequences as set forth in SEQ ID NO. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 and 31, or a functional fragment, derivate, allele, homolog or isoenzyme of the enzyme encoding amino acid sequence,
- a₃) is a partial nucleotide sequence which corresponds to a full length nucleotide sequence selected from the group consisting of sequences as set forth in SEQ ID NO. 5, 7, 9, 12, 25, 26, 28 or 31, or a portion, derivate, allele or homolog thereof;
- a_4) is a nucleotide sequence which is at least 40% identical to a nucleotide sequence selected from the group consisting of those sequences set forth in SEQ ID NO. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 and 31,
- b₁) is a gene construct comprising a nucleotide sequence a), operably linked to a heterologous nucleic acid, and
- c_1) is a vector comprising a gene construct b_1), or a nucleotide sequence a_1).
- 41. A transgenic cell or organism according to claim 40, which is an eucaryotic cell or organism.
- 42. A transgenic cell or organism according to claim 40, which is a yeast cell or a plant cell or a plant.
- 43. A transgenic cell or organism according to claim 40 having an altered biosynthetic pathway for the production of

triacylglycerol, characterized by the prevention of accumulation of undesirable fatty acids, which are harmful if present in high amounts in membrane lipids.

- 44. A transgenic cell or organism according to claim 40 having an altered, increased oil content.
- 45. A transgenic cell or organism according to claim 40, wherein the activity of PDAT is altered, characterized by an alteration in gene expression, catalytic activity and/or regulation of activity of the enzyme.
- 46. A process for the production of triacylglycerol, comprising growing a transgenic cell or organism according to claim 40 under conditions whereby the said nucleotide sequence
- a₁) is a nucleotide sequence as set forth in SEQ ID NO. 1 or a homologous nucleotide sequence which is at least about 40% identical to a nucleotide sequence of SEQ ID NO. 1,
- a₂) is a nucleotide sequence, a portion, derivate, allele or homolog thereof selected from the group consisting of sequences as set forth in SEQ ID NO. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 and 31, or a functional fragment, derivate, allele, homolog or isoenzyme of the enzyme encoding amino acid sequence,
- a₃) is a partial nucleotide sequence which corresponds to a full length nucleotide sequence selected from the group consisting of sequences as set forth in SEQ ID NO. 5, 7, 9, 12, 25, 26, 28 or 31, or a portion, derivate, allele or homolog thereof;
- a₄) is a nucleotide sequence which is at least 40% identical to a nucleotide sequence selected from the group consisting of those sequences set forth in SEQ ID NO. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 and 31,

is expressed.

- 47. Triacylglycerols produced by a process according to claim 46.
 - 48. Use of a nucleotide sequence selected from the group of
- a_1) is a nucleotide sequence as set forth in SEQ ID NO. 1 or a homologous nucleotide sequence which is at least about 40% identical to a nucleotide sequence of SEQ ID NO. 1,
- a₂) is a nucleotide sequence, a portion, derivate, allele or homolog thereof selected from the group consisting of sequences as set forth in SEQ ID NO. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 and 31, or a functional fragment, derivate, allele, homolog or isoenzyme of the enzyme encoding amino acid sequence,
- a₃) is a partial nucleotide sequence which corresponds to a full length nucleotide sequence selected from the group consisting of sequences as set forth in SEQ ID NO. 5, 7, 9, 12, 25, 26, 28 or 31, or a portion, derivate, allele or homolog thereof;
- a₄) is a nucleotide sequence which is at least 40% identical to a nucleotide sequence selected from the group consisting of those sequences set forth in SEQ ID NO. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 and 31,
- or an enzyme selected from the group of
- d₁) is an enzyme, designated as phospholipid:diacylglycerol acyltransferase (PDAT), catalyzing in an acyl-CoA-independent reaction the transfer of fatty acids from phospholipids to diacylglycerol in the biosynthetic pathway for the production of triacylglycerol and comprising an amino acid sequence as set forth in SEQ ID NO. 2 or a functional fragment, derivate, allele, homolog or isoenzyme thereof,
- $d_2)$ is an enzyme d), comprising an amino acid sequence as set forth in SEQ ID NO. 16, 20 or 22, or a functional fragment,

derivate, allele, homolog or isoenzyme thereof,

- d₃) is an enzyme d₁), comprising an amino acid sequence selected from the group consisting of sequences as set forth in SEQ ID NO. 6, 8, 13, 14, 15, 17, 18, 25 and 27, or a functional fragment, derivate, allele, homolog or isoenzyme thereof, for the production of triacylglycerol and/or triacylglycerols with uncommon fatty acids, comprising medium chain fatty acids, hydroxylated fatty acids, epoxygenated fatty acids and acetylenic fatty acids.
- 49. Use of a nucleotide sequence selected from the group of a₁) is a nucleotide sequence as set forth in SEQ ID NO. 1 or a homologous nucleotide sequence which is at least about 40% identical to a nucleotide sequence of SEQ ID NO. 1,
- a₂) is a nucleotide sequence, a portion, derivate, allele or homolog thereof selected from the group consisting of sequences as set forth in SEQ ID NO. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 and 31, or a functional fragment, derivate, allele, homolog or isoenzyme of the enzyme encoding amino acid sequence,
- a₃) is a partial nucleotide sequence which corresponds to a full length nucleotide sequence selected from the group consisting of sequences as set forth in SEQ ID NO. 5, 7, 9, 12, 25, 26, 28 or 31, or a portion, derivate, allele or homolog thereof;
- a₄) is a nucleotide sequence which is at least 40% identical to a nucleotide sequence selected from the group consisting of those sequences set forth in SEQ ID NO. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 and 31,
- or an enzyme selected from the group of
- d_1) is an enzyme, designated as phospholipid:diacylglycerol acyltransferase (PDAT), catalyzing in an acyl-CoA-independent

reaction the transfer of fatty acids from phospholipids to diacylglycerol in the biosynthetic pathway for the production of triacylglycerol and comprising an amino acid sequence as set forth in SEQ ID NO. 2 or a functional fragment, derivate, allele, homolog or isoenzyme thereof,

- d_2) is an enzyme d_1), comprising an amino acid sequence as set forth in SEQ ID NO. 16, 20 or 22, or a functional fragment, derivate, allele, homolog or isoenzyme thereof,
- d_3) is an enzyme d_1), comprising an amino acid sequence selected from the group consisting of sequences as set forth in SEQ ID NO. 6, 8, 13, 14, 15, 17, 18, 25 and 27, or a functional fragment, derivate, allele, homolog or isoenzyme thereof, for the transformation of any cell or organism in order to be expressed in this cell or organism and result in an altered, preferably increased oil content of this cell or organism.

REMARKS

The claims have been amended to eliminate multiple dependency and to place them in better form for U.S. practice. Further, amendments made in the international stage, albeit not under Article 19, are also incorporated. The changes made in the claims were as follows:

- i. the subject-matter of claim 2 and 3 was included in claim 1
 (now claim 28);
- ii. claim 29 was introduced according to the disclosed homologous nucleotide sequence on page 6 of the specification;
- iii. claims 7, 8 and 25 were deleted;
- iv. claims 30-42 essentially correspond to claims 4-6 and 9-18 of the international application;
- v. claim 23 was incorporated into claim 19, which is now claim 43;
- vi claim 20, now 44, was amended by defining the altered oil content according to page 9, line 9;
- vii. claim 22 was incorporated into claim 21, which is now claim 45;
- viii. claim 46 essentially corresponds to claim 24 of the international application.
- ix. in claim 26, now 47, triacylglycerols with uncommon fatty acids were defined according to page 10, line 29;

- x. claim 48 essentially corresponds to claim 27 of the international application;
- xi. in the other claims, editorial amendments were made.
 Favorable action on the application is solicited.

Respectfully submitted,

KEIL & WEINKAUF

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O 7 JUN 2002

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

的 re Application of

DAHLQUIST et al.

BOX PCT

Serial No. 09/937,779

Filed: September 28, 2001

NEW CLASS OF ENZYMES IN THE BIOSYNTHETIC PATHWAY FOR THE

PRODUCTION OF TRIACYCLOGLYCEROL AND RECOMBINANT DNA

MOLECULES ENCODING THESE ENZYMES

I hereby certify that this correspondence is being deposited with the United States Postal Service as first class mail in an envelope addressed to Commissioner of Patents and Trademarks, Washington, D.C. 20231, on:

June 5,

Karen Stamper

Signature

Date of Signature

Honorable Commissioner of Patents and Trademarks Washington, D.C. 20231

PRELIMINARY AMENDMENT

and

RESPONSE TO NOTIFICATION OF MISSING REQUIREMENTS UNDER 35 USC 371

Sir:

In response to the Notification of Missing Requirements under 35 USC 371, a copy of the Sequence Listing in computer readable form is attached hereto. The content of the paper copy of the Sequence Listing and the copy of the Sequence Listing in computer readable form is the same, and includes no new matter.

IN THE SPECIFICATION

Delete the sequence listing in the specification on pages 1/58 to 58/58 and substitute with the attached replacement sequence listing on separate pages 1-52.

REMARKS

It is believed that by submitting the present amendment and sequence listing diskette, the application now fully complies with the requirements of 37 CFR 1.821-1.825. Favorable action by the examiner is solicited.

Please charge any shortage in fees due in connection with the filing of this paper, including Extension of Time fees to Deposit Account No. 11-0345. Please credit any excess fees to such account.

Respectfully submitted,

KEIL & WEINKAUF

06/13/2002 HKAYPAGH 00000090 110345 09937779

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130.00 CH

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A NEW CLASS OF ENZYMES IN THE BIOSYNTHETIC PATHWAY FOR THE PRODUCTION OF TRIACYLGLYCEROL AND RECOMBINANT DNA MOLECULES ENCODING THESE ENZYMES

The present invention relates to the isolation, identification and characterization of recombinant DNA molecules encoding enzymes catalysing the transfer of fatty acids from phospholipids to diacylglycerol in the biosynthetic pathway for the production of triacylglycerol.

Triacylglycerol (TAG) is the most common lipid-based energy reserve in nature. The main pathway for synthesis of TAG is believed to involve three sequential acyl-transfers from acyl-CoA to a glycerol backbone (1, 2). For many years, acyl-CoA: diacylglycerol acyltransferase (DAGAT), which catalyzes the third acyl transfer reaction, was thought to be the only unique enzyme involved in TAG synthesis. It acts by diverting diacylglycerol (DAG) from membrane lipid synthesis into TAG (2). Genes encoding this enzyme were recently identified both in the mouse (3) and in plants (4, 5), and the encoded proteins were shown to be homologous to acyl-CoA: cholesterol acyltransferase (ACAT). It was also recently reported that another DAGAT exists in the oleaginous fungus *Mortierella ramanniana*, which is unrelated to the mouse DAGAT, the ACAT gene family or to any other known gene (6).

The instant invention relates to novel type of enzymes and their encoding genes for transformation. More specifically, the invention relates to use of a type of genes encoding a not previously described type of enzymes hereinafter designated phospholipid:diacylglycerol acyltransferases (PDAT), whereby this enzyme catalyses an acyl-CoA-independent reaction. The said type of genes expressed alone in transgenic organisms will enhance the total amount of oil (triacylglycerols) produced in the cells. The PDAT genes, in combination with a gene for the synthesis of an uncommon fatty acid will, when expressed in transgenic organisms, enhance the levels of the uncommon fatty acids in the triacylglycerols.

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There is considerable interest world-wide in producing chemical feedstock, such as fatty acids, for industrial use from renewable plant resources rather than non-renewable petrochemicals. This concept has broad appeal to manufacturers and consumers on the basis of resource conservation and provides significant opportunity to develop new industrial crops for agriculture.

There is a diverse array of unusual fatty acids in oils from wild plant species and these have been well characterised. Many of these acids have industrial potential and this has led to interest in domesticating relevant plant species to enable agricultural production of particular fatty acids.

Development in genetic engineering technologies combined with greater understanding of the biosynthesis of unusual fatty acids now makes it possible to transfer genes coding for key enzymes involved in the synthesis of a particular fatty acid from a wild species into domesticated oilseed crops. In this way individual fatty acids can be produced in high purity and quantities at moderate costs.

In all crops like rape, sunflower, oilpalm etc., the oil (i.e. triacylglycerols) is the most valuable product of the seeds or fruits and other compounds like starch, protein, and fibre is regarded as by-products with less value. Enhancing the quantity of oil per weight basis at the expense of other compounds in oil crops would therefore increase the value of crop. If genes, regulating the allocation of reduced carbon into the production of oil can be up-regulated, the cells will accumulate more oil on the expense of other products. Such genes might not only be used in already high oil producing cells, such as oil crops, but could also induce significant oil production in moderate or low oil containing crops such as e.g. soy, oat, maize, potato, sugarbeats, and turnips as well as in micro-organisms.

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Many of the unusual fatty acids of interest, e.g. medium chain fatty acids, hydroxy fatty acids, epoxy fatty acids and acetylenic fatty acids, have physical properties that are distinctly different from the common plant fatty acids. The present inventors have found that, in plant species naturally accumulating these uncommon fatty acids in their seed oil (i.e. triacylglycerol), these acids are absent, or present in very low amounts in the membrane (phospho)lipids of the seed. The low concentration of these acids in the membrane lipids is most likely a prerequisite for proper membrane function and thereby for proper cell functions. One aspect of the invention is that seeds of transgenic crops can be made to accumulate high amounts of uncommon fatty acids if these fatty acids are efficiently removed from the membrane lipids and channelled into seed triacylglycerols.

The inventors have identified a novel class of enzymes in plants catalysing the transfer of fatty acids from phospholipids to diacylglycerol in the production of triacylglycerol through an acyl-CoA-independent reaction and that these enzymes (phospholipid:diacylglycerol acyltransferases, abbreviated as PDAT) are involved in the removal of hydroxylated, epoxygenated fatty acids, and probably also other uncommon fatty acids such as medium chain fatty acids, from phospholipids in plants.

This enzyme reaction was shown to be present in microsomal preparations from baker's yeast (*Saccharomyces cerevisiae*). The instant invention further pertains to an enzyme comprising an amino acid sequence as set forth in SEQ ID No. 2 or a functional fragment, derivate, allele, homolog or isoenzyme thereof. A so called ,knock out' yeast mutant, disrupted in the respective gene was obtained and microsomal membranes from the mutant was shown to totally lack PDAT activity. Thus, it was proved that the disrupted gene encodes a PDAT enzyme (SEQ ID NO. 1 and 2). Furtherm, this PDAT enzyme is characterized through the amino acid sequence as set forth in SEQ ID NO 2 containing a lipase motif of the conserved sequence string FXKWVEA.

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The instant invention pertains further to an enzyme comprising an amino acid sequence as set forth in SEQ ID NO. 16, 20 or 22 or a functional fragment, derivate, allele, homolog or isoenzyme thereof.

Further genes and/or proteins of so far unknown function were identified and are contemplated within the scope of the instant invention. A gene from Schizosaccharomyces pombe, SPBC776.14 (SEQ ID. NO. 3), a putative open reading frame CAA22887 of the SPBC776.14 (SEQ ID NO. 13) were identified. Further Arabidopsis thaliana genomic sequences (SEQ ID NO. 4, 10 and 11) coding for putative proteins were identified, as well as a putative open reading frame AAC80628 from the A. thaliana locus AC 004557 (SEQ ID NO. 14) and a putative open reading frame AAD10668 from the A. thaliana locus AC 003027 (SEQ ID NO. 15) were identified.

Also, a partially sequenced cDNA clone from Neurospora crassa (SEQ ID NO. 9) and a Zea mays EST (Extended Sequence Tac) clone (SEQ ID NO. 7) and corresponding putative amino acid sequence (SEQ ID NO. 8) were identified. Finally, two cDNA clones were identified, one Arabidopsis thaliana EST (SEQ ID NO. 5 and corresponding predicted amino acid sequence SEQ ID NO. 6) and a Lycopersicon esculentum EST clone (SEQ ID NO. 12) were identified. Further, enzymes designated as PDAT comprising an amino acid sequence selected from the group consisting of sequences as set forth in SEQ ID NO 6, 17, 18, 25 or 27 containing a lipase motif FXKWVEA are contemplated within the scope of the invention. Moreover, an enzyme comprising an amino acid sequence encoded through a nucleotide sequence, a portion, derivate, allele or homolog thereof selected from the group consisting of sequences as set forth in SEQ ID No. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 or 31 or a functional fragment, derivate, allele, homolog or isoenzyme of the enzyme encoding amino acid sequence are included within the scope of the invention.

A functional fragment of the instant enzyme is understood to be any polypeptide sequence which shows specific enzyme activity of a phospholipid:diacylglycerol acyltransferase (PDAT). The length of the functional fragment can for example vary in a range from about 660 ± 10 amino acids to

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 660 ± 250 amino acids, preferably from about 660 ± 50 to 660 ± 100 amino acids, whereby the "basic number" of 660 amino acids corresponds in this case to the polypeptide chain of the PDAT enzyme of SEQ ID NO. 2 encoded by a nucleotide sequence according to SEQ ID NO. 1. Consequently, the "basic number" of functional fullength enzyme can vary in correspondance to the encoding nucleotide sequence.

A portion of the instant nucleotide sequence is meant to be any nucleotide sequence encoding a polypeptid which shows specific activity of a phospholipid:diacylglycerol acyltransferase (PDAT). The length of the nucleotide portion can vary in a wide range of about several hundreds of nucleotides based upon the coding region of the gene or a highly conserved sequence. For example the length varies in a range form about 1900 ± 10 to 1900 ± 1000 nucleotides, preferably form about 1900 ± 50 to 1900 ± 700 and more preferably form about 1900 ± 100 to 1900 ± 500 nucleotides, whereby the "basic number" of 1900 nucleotides corresponds in this case to the encoding nucleotide sequence of the PDAT enzyme of SEQ ID NO. 1. Consequently, the "basic number" of functional fullength gene can vary.

An allelic variant of the instant nucleotide sequence is understood to be any different nucleotide sequence which encodes a polypeptide with a functionally equivalent function. The alleles pertain naturally occurring variants of the instant nucleotide sequences as well as synthetic nucleotide sequences produced by methods known in the art. Contemplated are even altered nucleotide sequences which result in an enzyme with altered activity and/or regulation or which is resistant against specific inhibitors. The instant invention further includes natural or synthetic mutations of the originally isolated nucleotide sequences. These mutations can be substitution, addition, deletion, inversion or insertion of one or more nucleotides.

A homologous nucleotide sequence is understood to be a complementary sequence and/or a sequence which specifically hybridizes with the instant nucleotide sequence. Hybridizing sequences include similar sequences selected from the group of DNA or RNA which specifically interact to the instant

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nucleotide sequences under at least moderate stringency conditions which are known in the art. A preferred, non-limiting example of stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45°C, followed by one or more washes in 0.2 X SSC, 0.1% SDS at 50-65°C. This further includes short nucleotide sequences of e.g. 10 to 30 nucleotides, preferably 12 to 15 nucleotides. Included are also primer or hybridization probes.

A homologous nucleotide sequence included within the scope of the instant invention is a sequence which is at least about 40%, preferably at least about 50 % or 60%, and more preferably at least about 70%, 80% or 90% and most preferably at least about 95%, 96%, 97%, 98% or 99% or more homologous to a nucleotide sequence of SEQ ID NO. 1.

All of the aforementioned definitions are true for amino acid sequences and functional enzymes and can easily transferred by a person skilled in the art.

Isoenzymes are understood to be enzymes which have the same or a similar substrate specifity and/or catalytic activity but a different primary structure.

In a first embodiment, this invention is directed to nucleic acid sequences that encode a PDAT. This includes sequences that encode biologically active PDATs as well as sequences that are to be used as probes, vectors for transformation or cloning intermediates. The PDAT encoding sequence may encode a complete or partial sequence depending upon the intended use. All or a portion of the genomic sequence, cDNA sequence, precursor PDAT or mature PDAT is intended.

Further included is a nucleotide sequence selected from the group consisting of sequences set forth in SEQ ID No. 1, 3, 4, 10, 11, 19, 21, 23, 24, 29 or 30 or a portion, derivate, allele or homolog thereof. The invention pertains a partial nucleotide sequence corresponding to a fullength nucleotide sequence selected from the group consisting of sequences set forth in SEQ ID No. 5, 7, 9, 12, 25, 26, 28 or 31 or a portion, derivate, allele or homolog thereof. Moreover, a

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nucleotide sequence comprising a nucleotide sequence which is at least 40% homologous to a nucleotide sequence selected form the group consisting of those sequences set forth in SEQ ID No. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 or 31 is contemplated within the scope of the invention.

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The instant invention pertains to a gene construct comprising a said nucleotide sequences of the instant invention which is operably linked to a heterologous nucleic acid.

The term operably linked means a serial organisation e.g. of a promotor, coding sequence, terminator and/or further regulatory elements whereby each element can fulfill its original function during expression of the nucleotide sequence.

Further, a vector comprising of a said nucleotide sequence of the instant invention is contemplated in the instant invention. This includes also an expression vector as well as a vector further comprising a selectable marker gene and/or nucleotide sequences for the replication in a host cell and/or the integration into the genome of the host cell.

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In a different aspect, this invention relates to a method for producing a PDAT in a host cell or progeny thereof, including genetically engineered oil seeds, yeast and moulds or any other oil accumulating organism, via the expression of a construct in the cell. Cells containing a PDAT as a result of the production of the PDAT encoding sequence are also contemplated within the scope of the invention.

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Further, the invention pertains a transgenic cell or organism containing a said nucleotide sequence and/or a said gene construct and/or a said vector. The object of the instant invention is further a transgenic cell or organism which is an eucaryotic cell or organism. Preferably, the transgenic cell or organism is a yeast cell or a plant cell or a plant. The instant invention further pertains said transgenic cell or organism having an altered biosynthetic pathway for the production of triacylglycerol. A transgenic cell or organism having an altered oil content is also contemplated within the scope of this invention.

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Further, the invention pertains a transgenic cell or organism wherein the activity of PDAT is altered in said cell or organism. This altered activity of PDAT is characterized by an alteration in gene expression, catalytic activity and/or regulation of activity of the enzyme. Moreover, a transgenic cell or organism is included in the instant invention, wherein the altered biosynthetic pathway for the production of triacylglycerol is characterized by the prevention of accumulation of undesirable fatty acids in the membrane lipids.

In a different embodiment, this invention also relates to methods of using a DNA sequence encoding a PDAT for increasing the oil-content within a cell.

Another aspect of the invention relates to the accommodation of high amounts of uncomman fatty acids in the triacylglycerol produced within a cell, by introducing a DNA sequence producing a PDAT that specifically removes these fatty acids from the membrane lipids of the cell and channel them into triacylglycerol. Plant cells having such a modification are also contemplated herein.

Further, the invention pertains a process for the production of triacylglycerol, comprising growing a said transgenic cell or organism under conditions whereby the said nucleotide sequence is expressed and whereby the said transgenic cells comprising a said enzyme catalysing the transfer of fatty acids from phospholipids to diacylglycerol forming triacylglycerol.

Moreover, triacylglycerols produced by the aforementioned process are included in scope of the instant invention.

Object of the instant invention is further the use of an instant nucleotide sequence and/or a said enzyme for the production of triacylglycerol and/or triacylglycerols with uncommon fatty acids. The use of a said instant nucleotide sequence and/or a said enzyme of the instant invention for the transformation of any cell or organism in order to be expressed in this cell or organism and

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result in an altered, preferably increased oil content of this cell or organism is also contemplated within the scope of the instant invention.

A PDAT of this invention includes any sequence of amino acids, such as a protein, polypeptide or peptide fragment obtainable from a microorganism, animal or plant source that demonstrates the ability to catalyse the production of triacylglycerol from a phospholipid and diacylglycerol under enzyme reactive conditions. By "enzyme reactive conditions" is meant that any necessary conditions are available in an environment (e.g., such factors as temperature, pH, lack of inhibiting substances) which will permit the enzyme to function.

Other PDATs are obtainable from the specific sequences provided herein. Furthermore, it will be apparent that one can obtain natural and synthetic PDATs, including modified amino acid sequences and starting materials for synthetic-protein modelling from the examplified PDATs and from PDATs which are obtained through the use of such examplified sequences. Modified amino acid sequences include sequences that have been mutated, truncated, increased and the like, whether such sequences were partially or wholly synthesised. Sequences that are actually purified from plant preparations or are identical or encode identical proteins thereto, regardless of the method used to obtain the protein or sequence, are equally considered naturally derived.

Further, the nucleic acid probes (DNA and RNA) of the present invention can be used to screen and recover "homologous" or "related" PDATs from a variety of plant and microbial sources.

Further, it is also apparent that a person skilled in the art can, with the information provided in this application, in any organism identify a PDAT activity, purify an enzyme with this activity and thereby identify a "non-homologous" nucleic acid sequence encoding such an enzyme.

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The present invention can be essentially characterized by the following aspects:

- 1. Use of a PDAT gene (genomic clone or cDNA) for transformation.
- Use of a DNA molecule according to item 1 wherein said DNA is used for transformation of any organism in order to be expressed in this organism and result in an active recombinant PDAT enzyme in order to increase oil content of the organism.
 - Use of a DNA molecule of item 1 wherein said DNA is used for transformation of any organism in order to prevent the accumulation of undesirable fatty acids in the membrane lipids.
 - 4. Use according to item 1, wherein said PDAT gene is used for transforming transgenic oil accumulating organisms engineered to produce any uncommon fatty acid which is harmful if present in high amounts in membrane lipids, such as medium chain fatty acids, hydroxylated fatty acids, epoxygenated fatty acids and acetylenic fatty acids.
 - 5. Use according to item 1, wherein said PDAT gene is used for transforming organisms, and wherein said organisms are crossed with other oil accummulating organisms engineered to produce any uncommon fatty acid which is harmful if present in high amounts in membrane lipids, comprising medium chain fatty acids, hydroxylated fatty acids, epoxygenated fatty acids and acetylenic fatty acids.
 - 6. Use according to item 1, wherein the enzyme encoded by said PDAT gene or cDNA is coding for a PDAT with distinct acyl specificity.
- 7. Use according to item 1 wherein said PDAT encoding gene or cDNA, is derived from Saccharornyces cereviseae, or contain nucleotide sequences coding for an amino acid sequence 30% or more identical to the amino acid sequence of PDAT as presented in SEQ. ID. NO. 2.
- 8. Use according to item 1 wherein said PDAT encoding gene or cDNA is derived from *Saccharornyces cereviseae*, or contain nucleotide sequences coding for an amino acid sequence 40% or more *identical* to the amino acid sequence of PDAT as presented in SEQ. ID. NO. 2.

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- 9. Use according to item 1 wherein said PDAT encoding gene or cDNA is derived from *Saccharornyces cereviseae*, or contain nucleotide sequences coding for an amino acid sequence 60% or more *identical* to the amino acid sequence of PDAT as presented in SEQ. ID. NO. 2.
- 10. Use according to item 1 wherein said PDAT encoding gene or cDNA is derived from *Saccharornyces cereviseae*, or contain nucleotide sequences coding for an amino acid sequence 80% or more identical to the amino acid sequence of PDAT as presented in SEQ. ID. NO. 2.
 - 11. Use according to item 1 wherein said PDAT encoding gene or cDNA is derived from plants or contain nucleotide sequences coding for an amino acid sequence 40% or more identical to the amino acid sequence of PDAT from *Arabidopsis thaliana* or to the protein encoded by the fullength counterpart of the partial Zea mays, Lycopericon esculentum, or Neurospora crassa cDNA clones.
- 12. Transgenic oil accumulating organisms comprising, in their genome, a PDAT gene transferred by recombinant DNA technology or somatic hybridization.
 - 13. Transgenic oil accumulating organisms according to item 12 comprising, in their genome, a PDAT gene having specificity for substrates with a particular uncommon fatty acid and the gene for said uncommon fatty acid.
 - 14. Transgenic organisms according to item 12 or 13 which are selected from the group consisting of fungi, plants and animals.
 - 15. Transgenic organisms according to item 12 or 13 which are selected from the group of agricultural plants.
- 16. Transgenic organisms according to item 12 or 13 which are selected from the group of agricultural plants and where said PDAT gene is expressed under the control of a storage organ specific promotor.
 - 17. Transgenic organisms according to item 12 or 13 which are selected from the group of agricultural plants and where said PDAT gene is expressed under the control of a seed promotor.
 - 18. Oils from organisms according to item 12 17.
 - 19. A method for altering acyl specificity of a PDAT by alteration of the nucleotide sequence of a naturally occurring encoding gene and as a

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consequence of this alternation creating a gene encoding for an enzyme with novel acyl specifity.

- 20. A protein encoded by a DNA molecule according to item 1 or a functional fragment thereof.
- 21. A protein of item 20 designated phospholipid:diacylglycerol acyltransferase.
 - 22. A protein of item 21 which has a distinct acyl specificity.
 - 23. A protein of item 13 having the amino acid sequence as set forth in SEQ, ID NO. 2, 13, 14 or 15 (and the proteins encoded by the fullength or partial genes set forth in SEQ. ID. NO. 1, 3, 4, 5, 7, 9, 10, 11 or 12) or an amino acid sequence with at least 30 % homology to said amino acid sequence.
 - 24. A protein of item 23 isolated from Saccharomyces cereviseae.

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20 General methods:

Yeast strains and plasmids. The wild type yeast strains used were either FY1679 (MATα his3-Δ200 leu2-Δ1 trp1-Δ6 ura3-52) or W303-1A (MATa ADE2-1 can1-100 his3-11,15 leu2-3,112 trp1-1 ura3-1) (7). The YNR008w::KanMX2 disruption strain FVKT004-04C(AL), which is congenic to FY1679, was obtained from the Euroscarf collection (8). A 2751 bp fragment containing the YNR008w gene with 583 bp of 5' and 183 bp of 3' flanking DNA was amplified W303-1A genomic DNA using Tag polymerase from with TCTCCATCTTCTGCAAAACCT-3' and 5'-CCTGTCAAAAACCTTCTCCTC-3' as primers. The resulting PCR product was purified by agarose gel electrophoresis and cloned into the EcoRV site of pBluescript (pbluescript-pdat). For complementation experiments, the cloned fragment was released from pBluescript by HindIII-Sacl digestion and then cloned between the HindIII and SacI sites of pFL39 (9), thus generating pUS1. For overexpression of the PDAT

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gene, a 2202 bp *Eco*RI fragment from the pBluscript plasmid which contains only 24 bp of 5' flanking DNA was cloned into the BamHI site of the *GAL1-TPK2* expression vector pJN92 (12), thus generating pUS4.

Microsomal preparations. Microsomes from developing seeds of sunflower (Helianthus annuus), Ricinus communis and Crepis palaestina were prepared using the procedure of Stobart and Stymne (11). To obtain yeast microsomes, 1g of yeast cells (fresh weight) was re-suspended in 8 ml of ice-cold buffer (20 mM Tris-Cl, pH 7.9, 10 mM MgCl₂, 1 mM EDTA. 5 % (v/v) glycerol, 1 mM DTT, 0.3 M ammonium sulfate) in a 12 ml glass tube. To this tube, 4 ml of glass beads (diameter 0.45-0.5 mm) were added, and the tube was then heavily shaken (3 x 60 s) in an MSK cell homogenizer (B. Braun Melsungen AG, Germany). The homogenized suspension was centrifuged at 20,000 x g for 15 min at 6°C and the resulting supernatant was again centrifuged at 100,000 x g for 2 h at 6°C. The 100,000 x g pellet was resuspended in 0.1 M potassium phosphate (pH 7.2), and stored at -80°C. It is subsequently referred to as the crude yeast microsomal fraction.

Lipid substrates. Radio-labeled ricinoleic (12-hydroxy-9-octadecenoic) and vernolic (12,13-epoxy-9-octadecenoic) acids were synthesized enzymatically from [1-¹⁴C]oleic acid and [1-¹⁴C]linoleic acid, respectively, by incubation with microsomal preparations from seeds of *Ricinus communis* and *Crepis palaestina*, respectively (12). The synthesis of phosphatidylcholines (PC) or phosphatidylethanolamines (PE) with ¹⁴C-labeled acyl groups in the *sn-*2 position was performed using either enzymatic (13), or synthetic (14) acylation of [¹⁴C]oleic, [¹⁴C]ricinoleic, or [¹⁴C]vernolic acid. Dioleoyl-PC that was labeled in the *sn-*1 position was synthesized from *sn-*1-[¹⁴C]oleoyl-lyso-PC and unlabeled oleic acid as described in (14). *Sn-*1-oleoyl-*sn-*2-[¹⁴C]ricinoleoyl-DAG was synthesized from PC by the action of phospholipase C type XI from *B. Cereus* (Sigma Chemical Co.) as described in (15). Monovernoloyl- and divernoleoyl-DAG were synthesized from TAG extracted from seeds of *Euphorbia lagascae*, using the TAG-lipase (Rizhopus arrhizus, Sigma Chemical

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Co.) as previously described (16). Monoricinoleoyl-TAG was synthesized according to the same method using TAG extracted from Castor bean.

Lipid analysis. Total lipid composition of yeast were determined from cells harvested from a 40 ml liquid culture, broken in a glass-bead shaker and extracted into chloroform as described by Bligh and Dyer (17), and then separated by thin layer chromatography in hexane/diethylether/acetic acid (80:20:1) using pre-coated silica gel 60 plates (Merck). The lipid areas were located by brief exposure to I2 vapors and identified by means of appropriate standards. Polar lipids, sterol-esters and triacylglycerols, as well as the remaining minor lipid classes, referred to as other lipids, were excised from the plates. Fatty acid methylesters were prepared by heating the dry excised material at 85 °C for 60 min in 2% (v/v) sulfuric acid in dry methanol. The methyl esters were extracted with hexane and analyzed by GLC through a 50 m mm CP-Wax58-CB fused-silica column (Chrompack), with methylheptadecanoic acid as an internal standard. The fatty acid content of each fraction was quantified and used to calculate the relative amount of each lipid class. In order to determine the total lipid content, 3 ml aliquots from yeast cultures were harvested by centrifugation and the resulting pellets were washed with distilled water and lyophilized. The weight of the dried cells was determined and the fatty acid content was quantified by GLC-analyses after conversion to methylesters as described above. The lipid content was then calculated as nmol fatty acid (FA) per mg dry weight yeast.

Enzyme assays. Aliquots of crude microsomal fractions (corresponding to 10 nmol of microsomal PC) from developing plant seeds or yeast cells were lyophilized over night. ¹⁴C-Labeled substrate lipids dissolved in benzene were then added to the dried microsomes. The benzene was evaporated under a stream of N₂, leaving the lipids in direct contact with the membranes, and 0.1 ml of 50 mM potassium phosphate (pH 7.2) was added. The suspension was thoroughly mixed and incubated at 30°C for the time period indicated, up to 90 min. Lipids were extracted from the reaction mixture using chloroform and separated by thin layer chromatography in hexane/diethylether/acetic acid

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(35:70:1.5) using silica gel 60 plates (Merck). The radioactive lipids were visualized and quantified on the plates by electronic autoradiography (Instant Imager, Packard, US).

<u>Yeast cultivation.</u> Yeast cells were grown at 28°C on a rotatory shaker in liquid YPD medium (1% yeast extract, 2% peptone, 2% glucose), synthetic medium (18) containing 2% (v/v) glycerol and 2% (v/v) ethanol, or minimal medium (19) containing 16 g/l of glycerol.

The instant invention is further characterized by the following examples which are not limiting:

Acyl-CoA-independent synthesis of TAG by oil seed microsomes. A large number of unusual fatty acids can be found in oil seeds (20). Many of these fatty acids, such as ricinoleic (21) and vernolic acids (22), are synthesized using phosphatidylcholin (PC) with oleoyl or linoleoyl groups esterified to the sn-2 position, respectively, as the immediate precursor. However, even though PC can be a substrate for unusual fatty acid synthesis and is the major membrane lipids in seeds, unusual fatty acids are rarely found in the membranes. Instead, they are mainly incorporated into the TAG. A mechanism for efficient and selective transfer of these unusual acyl groups from PC into TAG must therefore exist in oil seeds that accumulate such unusual fatty acids. This transfer reaction was biochemically characterized in seeds from castor bean (Ricinus communis) and Crepis palaestina, plants which accumulate high levels of ricinoleic and vernolic acid, respectively, and sunflower (Helianthus annuus), a plant which has only common fatty acids in its seed oil. Crude microsomal fractions from developing seeds were incubated with PC having ¹⁴C-labeled oleoyl, ricinoleoyl or vernoloyl groups at the *sn-2* position. After the incubation, lipids were extracted and analyzed by thin layer chromatography. We found that the amount of radioactivity that was incorporated into the neutral lipid fraction increased linearly over a period of 4 hours (data not shown). The distribution of [14C]acyl groups within the neutral lipid fraction was analyzed after 80 min (Fig. 1). Interestingly the amount and distribution of radioactivity

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between diffferent neutral lipids were strongly dependent both on the plant species and on the type of [14C]acyl chain. Thus, sunflower microsomes incorporated most of the label into DAG, regardless of the type of [14C]acyl group. In contrast, *R. communis* microsomes preferentially incorporated [14C]ricinoleoyl and [14C]vernoloyl groups into TAG, while [14C]oleyl groups mostly were found in DAG. *C. palaestina* microsomes, finally, incorporated only [14C]vernolyol groups into TAG, with [14C]ricinoleyl groups being found mostly as free fatty acids, and [14C]oleyl groups in DAG. This shows that the high *in vivo* levels of ricinoleic acid and vernolic acid in the TAG pool of *R. communis* and *C. palaestina*, respectively, can be explained by an efficient and selective transfer of the corresponding acyl groups from PC to TAG in these organisms.

The in-vitro synthesis of triacylglycerols in microsomal preparations of developing castor bean is summarized in table 1.

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PDAT: a novel enzyme that catalyzes acyl-CoA independent synthesis of TAG. It was investigated if DAG could serve both as an acyl donor as well as an acyl acceptor in the reactions catalyzed by the oil seed microsomes. Therefore, unlabeled divernoloyl-DAG was incubated with either sn-1-oleoyl-sn-2-[14C]ricinoleoyl-DAG or sn-1-oleoyl-sn-2-[14C]ricinoleoyl-PC in the presence of R. communis microsomes. The synthesis of TAG molecules containing both [14C]ricinoleoyl and vernoloyl groups was 5 fold higher when [14C]ricinoleoyl-PC served as acyl donor as compared to [14C]ricinoleoyl-DAG (fig.1B). These data strongly suggests that PC is the immediate acyl donor and DAG the acyl acceptor in the acyl-CoA-independent formation of TAG by oil seed microsomes. Therefore, this reaction is catalyzed by a new enzyme which we call phospholipid: diacylglycerol acyltransferase (PDAT).

<u>PDAT activity in yeast microsomes.</u> Wild type yeast cells were cultivated under conditions where TAG synthesis is induced. Microsomal membranes were prepared from these cells and incubated with *sn*-2-[¹⁴C]-ricinoleoyl-PC and DAG and the ¹⁴C-labeled products formed were analyzed. The PC-derived [¹⁴Clricinoleoyl groups within the neutral lipid fraction mainly were found in free

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fatty acids or TAG, and also that the amount of TAG synthesized was dependent on the amount of DAG that was added to the reaction (Fig.2). The *in vitro* synthesis of TAG containing both ricinoleoyl and vernoloyl groups, a TAG species not present *in vivo*, from exogenous added *sn*-2-[¹⁴C]ricinoleoyl-PC and unlabelled vernoloyl-DAG (Fig. 2, lane 3) clearly demonstrates the existence of an acyl-CoA-independent synthesis of TAG involving PC and DAG as substrates in yeast microsomal membranes. Consequently, TAG synthesis in yeast can be catalyzed by an enzyme similar to the PDAT found in plants.

The PDAT encoding gene in yeast.

A gene in the yeast genome (YNR008w) is known, but nothing is known about the function of YNR008w, except that the gene is not essential for growth under normal circumstances. Microsomal membranes were prepared from the yeast strain FVKT004-04C(AL) (8) in which this gene with unknown function had been disrupted. PDAT activity in the microsomes were assayed using PC with radiolabelled fatty acids at the sn-2 position. The activity was found to be completely absent in the disruption strain (Fig. 2 lane 4). Significantly, the activity could be partially restored by the presence of YNR008w on the single 2 lane 5). Moreover, acyl groups of copy plasmid pUS1 (Fig. phosphatidylethanolamine (PE) were efficiently incorporated into TAG by microsomes from the wild type strain whereas no incorporation occured from this substrate in the mutant strain (data not shown). This shows that YNR008w encodes a yeast PDAT which catalyzes the transfer of an acyl group from the sn-2 position of phospholipids to DAG, thus forming TAG. It should be noted that no cholesterol esters were formed from radioactive PC even in incubations with added ergosterols, nor were the amount of radioactive free fatty acids formed from PC affected by disruption of the YNR008w gene (data not shown). This demonstrates that yeast PDAT do not have cholesterol ester synthesising or phospholipase activities.

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Increased TAG content in yeast cells that overexpress PDAT. The effect of overexpressing the PDAT-encoding gene was studied by transforming a wild type yeast strain with the pUS4 plasmid in which the gene is expressed from

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the galactose-induced GAL1:TPK2 promoter. Cells containing the empty expression vector were used as a control. The cells were grown in synthetic glycerol-ethanol medium, and expression of the gene was induced after either 2 hours (early log phase) or 25 hours (stationary phase) by the addition of galactose. The cells were then incubated for another 21 hours, after which they were harvested and assays were performed. We found that overexpression of PDAT had no significant effect on the growth rate as determined by the optical density. However, the total lipid content, measured as µmol fatty acids per mg yeast dry weight, was 47% (log phase) or 29% (stationary phase) higher in the PDAT overexpressing strain than in the control. Furthermore, the polar lipid and sterolester content was unaffected by overexpression of PDAT. Instead, the elevated lipid content in these cells is entirely due to an increased TAG content (Fig. 3A,B). Thus, the amount of TAG was increased by 2-fold in PDAT overexpressing early log phase cells and by 40% in stationary phase cells. It is interesting to note that a significant increase in the TAG content was achieved by overexpressing PDAT even under conditions (i.e. in stationary phase) where DAGAT is induced and thus contributes significantly to TAG synthesis. In vitro PDAT activity assayed in microsomes from the PDAT overexpressing strain was 7-fold higher than in the control strain, a finding which is consistent with the increased levels of TAG that we observed in vivo (Fig. 3C). These results clearly demonstrate the potential use of the PDAT gene in increasing the oil content in transgenic organisms.

Substrate specificity of yeast PDAT. The substrate specificity of yeast PDAT was analyzed using microsomes prepared from the PDAT overexpressing strain (see Fig. 4). The rate of TAG synthesis, under conditions given in figure 4 with di-oleoyl-PC as the acyl-donor, was 0.15 nmol per min and mg protein. With both oleoyl groups of PC labeled it was possible, under the given assay conditions, to detect the transfer of 11 pmol/min of [14C]oleoyl chain into TAG and the formation of 15 pmol/min of lyso-PC. In microsomes from the PDAT-deficient strain, no TAG at all and only trace amounts of lyso-PC was detected, strongly suggesting that yeast PDAT catalyses the formation of equimolar amounts of TAG and lyso-PC when supplied with PC and DAG as

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substrates. The fact that somewhat more lyso-PC than TAG is formed can be explained by the presence of a phospholipase in yeast microsomes, which produces lyso-PC and unesterified fatty acids from PC.

The specificity of yeast PDAT for different acyl group positions was investigated by incubating the microsomes with di-oleoyl-PC carrying a [14Clacyl group either at the sn-1 position (Fig. 4A bar 2) or the sn-2 position (Fig. 4A bar 3). We found that the major ¹⁴C-labeled product formed in the former case was lyso-PC, and in the latter case TAG. We conclude that yeast PDAT has a specificity for the transfer of acyl groups from the sn-2 position of the phospholipid to DAG, thus forming sn-1-lyso-PC and TAG. Under the given assay conditions, trace amounts of 14C-labelled DAG is formed from the sn-1 labeled PC by the reversible action of a CDP-choline : choline phosphotransferase. This labeled DAG can then be further converted into TAG by the PDAT activity. It is therefore not possible to distinguish whether the minor amounts of labeled TAG that is formed in the presence of di-oleoyl-PC carrying a [14C]acyl group in the sn-1 position, is synthesized directly from the sn-1-labeled PC by a PDAT that also can act on the sn-1 postion, or if it is first converted to sn-1-labeled DAG and then acylated by a PDAT with strict selectivity for the transfer of acyl groups at the sn-2 position of PC. Taken together, this shows that the PDAT encoded by YNR008w catalyses an acyl transfer from the sn-2 position of PC to DAG, thus causing the formation of TAG and lyso-PC.

The substrate specificity of yeast PDAT was further analyzed with respect to the headgroup of the acyl donor, the acyl group transferred and the acyl chains of the acceptor DAG molecule. The two major membrane lipids of *S. cerevisiae* are PC and PE, and as shown in Fig. 4B (bars 1 and 2). dioleoyl-PE is nearly 4-fold more efficient than dioleoyl-PC as acyl donor in the PDAT-catalyzed reaction. Moreover, the rate of acyl transfer is strongly dependent on the type of acyl group that is transferred. Thus, a ricinoleoyl group at the *sn*-2 position of PC is 2.5 times more efficiently transferred into TAG than an oleoyl group in the same position (Fig. 4B bars 1 and 3). In contrast, yeast PDAT has

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no preference for the transfer of vernoloyl groups over oleoyl groups (Fig. 4B bars 1 and 4). The acyl chain of the acceptor DAG molecule also affects the efficiency of the reaction. Thus, DAG with a ricinoleoyl or a vernoloyl group is a more efficient acyl acceptor than dioleoyl-DAG (Fig. 4B bars 1, 5 and 6). Taken together, these results clearly show that the efficiency of the PDAT-catalyzed acyl transfer is strongly dependent on the properties of the substrate lipids.

PDAT genes. Nucleotide and amino acid sequences of several PDAT genes are given as SEQ ID No. 1 through 15. Futher provisional and/or partial sequences are given as SEQ ID NO 16 through 20 and 21 through 31, respectively. One of the Arabidopsis genomic sequences (SEQ ID NO. 4) identified an Arabidopsis EST cDNA clone; T04806. This cDNA clone was fully characterised and the nucleotide sequence is given as SEQ ID NO. 5. Based on the sequence homology of the T04806 cDNA and the Arabidopsis thaliana genomic DNA sequence (SEQ ID NO 4) it is apparent that an additional A is present at position 417 in the cDNA clone (data not shown). Excluding this nucleotide would give the amino acid sequence depicted in SEQ ID NO. 12.

Increased TAG content in seeds of Arabidopsis thaliana that express the yeast PDAT. For the expression of the yeast PDAT gene in Arabidopsis thaliana an EcoRI fragment from the pBluescript-PDAT was cloned together with napin promotor (25) into the vector pGPTV-KAN (26). A plasmid (pGNapPDAT) having the yeast PDAT gene in the correct orientation was identified and transformed into Agrobacterium tumefaciens. These bacteria were used to transform Arabidopsis thaliana columbia (C-24) plants using the root transformation method (27). Plants transformed with an empty vector were used as controls.

First generation seeds (T1) were harvested and germinated on kanamycin containing medium. Second generation seeds (T2) were pooled from individual plants and their fatty acid contents analysed by quantification of their methyl esthers by gas liquid chromatography after methylation of the seeds with 2% sulphuric acid in methanol at 85 °C for 1,5 hours. Quaritification was done with heptadecanoic acid methyl esters as internal standard.

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From the transformation with pGNapPDAT one T1 plant (26-14) gave raise to seven T2 plants of which 3 plants yielded seeds with statistically (in a mean difference two-sided test) higher oil content than seeds from T2 plants generated from T1 plant 32-4 transformed with an empty vector (table 2).

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Description of Figures

FIG. 1.

Metabolism of 14C-labeled PC into the neutral lipid fraction by plant microsomes. (A) Microsomes from developing seeds of sunflower, R. communis and C. palaestina were incubated for 80 min at 30°C with PC (8 nmol) having oleic acid in its sn-1 position, and either ¹⁴C-labeled oleic. ricinoleic or vernolic acid in its sn-2 position. Radioactivity incorporated in TAG (open bars), DAG (solid bars), and unsterified fatty acids (hatched bars) was quantified using thin layer chromatography followed by autoradiography, and is shown as percentage of added labeled substrate. (B) Synthesis in vitro of TAG carrying two vernoloyl and one [14C]ricinoleoyl group by microsomes from R. communis. The substrates added were unlabeled divernoloyl-DAG (5 nmol), together with either sn-1-oleoyl-sn-2-[14C]ricinoleoyl-DAG (0.4 nmol, 7700 dpm/nmol) or sn-1-oleoyl-sn-2-[14C]ricinoleoyl-PC (0.4 nmol, 7700 dpm/nmol). The microsomes were incubated with the substrates for 30 min at 30°C, after which samples were removed for lipid analysis as described in the section "general methods". The data shown are the average of two experiments.

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FIG. 2.

PDAT activity in yeast microsomes, as visualized by autoradiogram of neutral lipid products separated on TLC. Microsomal membranes (10 nmol of PC) from the wild type yeast strain FY1679 (lanes 1-3), a congenic yeast strain (FVKT004-04C(AL)) that is disrupted for YNR008w (lane 4) or the same disruption strain transformed with the plasmid pUS1, containing the YNR008w gene behind its native promotor (lane 5), were assayed for PDAT activity. As substrates, we used 2 nmol *sn*-1-oleoyl-*sn*-2-[¹⁴C]ricinoleoyl-PC together with either 5 nmol of dioleoyl-DAG (lanes 2, 4 and 5) or *rac*-oleoyl-vernoleoyl-DAG (lane 3). The enzymatic assay and lipid analysis was performed as described in Materials and Methods. The cells were precultured for 20 h in liquid YPD medium, harvested and re-suspended in an equal volume of minimal medium (19) containing 16 g/l glycerol. The cells were then grown for an additional 24 h

prior to being harvested. Selection for the plasmid was maintained by growing the transformed cells in synthetic medium lacking uracil (18). Abbreviations: 1-OH-TAG, monoricinoleoyl-TAG; 1-OH-1-ep-TAG, monoricinoleoyl-monovernoloyl-TAG; OH-FA, unesterified ricinoleic acid.

Fig. 3.

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Lipid content (A,B) and PDAT activity (C) in PDAT overexpressing yeast cells. The PDAT gene in the plasmid pUS4 was overexpressed from the galactoseinduced GAL1-TPK2 promotor in the wild type strain W303-1A (7). Its expression was induced after (A) 2 hours or (B) 25 hours of growth by the addition of 2% final concentration (w/v) of galactose. The cells were then incubated for another 22 hours before being harvested. The amount of lipids of the harvested cells was determined by GLC-analysis of its fatty acid contents and is presented as µmol fatty acids per mg dry weight in either TAG (open bar), polar lipids (hatched bar), sterol esters (solid bar) and other lipids (striped bar). The data shown are the mean values of results with three independent yeast cultures. (C) In vitro synthesis of TAG by microsomes prepared from yeast cells containing either the empty vector (vector) or the PDAT plasmid (+ PDAT). The cells were grown as in Fig. 3A. The substrate lipids dioleoyl-DAG (2.5 nmol) and sn-1-oleoyl-sn-2-[14C]-oleoyl-PC (2 nmol) were added to aliquots of microsomes (10 nmol PC), which were then incubated for 10 min at 28 °C. The amount of label incorporated into TAG was quantified by electronic autoradiography. The results shown are the mean values of two experiments.

25 FIG. 4.

Substrate specificity of yeast PDAT. The PDAT activity was assayed by incubating aliquots of lyophilized microsomes (10 nmol PC) with substrate lipids at 30°C for 10 min (panel A) or 90 min (panel B). Unlabeled DAG (2.5 nmol) was used as substrates together with different labeled phospholipids, as shown in the figure. (A) *Sn*-position specificity of yeast PDAT regarding the acyl donor substrate. Dioleoyl-DAG together with either *sn*-1-[¹⁴C]oleoyl-*sn*-2-[¹⁴C]oleoyl-PC (*sn*1-[¹⁴C]-PC) or *sn*-1-oleoyl-sn-2-[¹⁴C]oleoyl-PC (*sn*2-[¹⁴C]-PC). (B) Specificity of yeast PDAT regarding

phospholipid headgroup and of the acyl composition of the phospholipid as well as of the diacylglycerol. Dioleoyl-DAG together with either sn-1-oleoyl-sn-2-[14C]oleoyl-PC (oleoyl-PC), sn-1-oleoyl-sn-2-[14C]oleoyl-PE (oleoyl-PE), sn-1oleoyl-sn-2-[14C]ricinoleoyl-PC (ricinoleoyl-PC) or sn-1-oleoyl-sn-2-[14C]vernoloyl-PC (vernoloyl-PC). In the experiments presented in the 2 bars to the far right, monoricinoleoyl-DAG (ricinoleoyl-DAG or mono-vernoloyl-DAG (vernoloyl-DAG) were used together with sn-1-oleoyl-sn-2-114Cl-oleoyl-PC. The label that was incorporated into TAG (solid bars) and lyso-PC (LPC, open bars) was quantified by electronic autoradiography. The results shown are the mean values of two experiments. The microsomes used were from W303-1A cells overexpressing the PDAT gene from the GAL1-TPK2 promotor, as described in Fig. 3. The expression was induced at early stationary phase and the cells were harvested after an additional 24 h.

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TAB.1:

In vitro synthesis of triacylglycerols in microsomal preparations of developing castor bean. Aliquots of microsomes (20 nmol PC) were lyophilised and substrate lipids were added in benzene solution: (A) 0.4 nmol [14C]-DAG (7760 dpm/nmol) and where indicated 1.6 nmol unlabelled DAG; (B) 0.4 nmol [14C]-DAG (7760 dpm/nmol) and 5 nmol unlabelled di-ricinoleoyl-PC and (C) 0.25 nmol [14C]-PC (4000 dpm/nmol) and 5 nmol unlabelled DAG. The benzene was evaporated by N2 and 0.1 ml of 50 mM potassium phosphate was added, thoroughly mixed and incubated at 30 °C for (A) 20 min.; (B) and (C) 30 min.. Assays were terminated by extraction of the lipids in chloroform. The lipids were then separated by thin layer chromatography on silica gel 60 plates (Merck; Darmstadt, Germany) in hexan/diethylether/acetic 35:70:1.5. The radioactive lipids were visualised and the radioactivity quantified on the plate by electronic autoradiography (Instant Imager. Packard, US). Results are presented as mean values of two experiments.

Radioactivity in different triacylglycerols (TAG) species formed. Abbreviations used: 1-OH-, mono-ricinoleoyl-; 2-OH, di-ricinoleoyl-; 3-OH-, triricinoleoyl; 1-

OH-1-ver-, mono-ricinoleoly-monovernoleoyl-; 1-OH-2-ver-, mono-ricinoleoyl-divernoleoyl-. Radiolabelled DAG and PC were prepared enzymatically. The radiolabelled ricinoleoyl group is attached at the sn-2-position of the lipid and unlabelled oleoyl group at the sn-1-position. Unlabelled DAG with vernoleoyl- or ricinoleoyl chains were prepared by the action of TAG lipase (6) on oil of Euphorbia lagascae or Castor bean, respectively. Synthetic di-ricinoleoyl-PC was kindly provided from Metapontum Agribios (Italy).

10 <u>TAB.2:</u>

Total fatty acids per mg of T2 seeds pooled from individual *Arabidopsis thaliana* plants transformed with yeast PDAT gene under the control of napin promotor (26-14) or transformed with empty vector (32-4).

 * = stastistical difference between control plants and PDAT transformed plants in a mean difference two-sided test at $\alpha = 5$.

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Description of the SEQ ID:

- SEQ ID NO. 1: Genomic DNA sequence and suggested amino acid sequence of the Saccharomyces cerevisiae PDAT gene, YNR008w, with GenBank accession number Z71623 and Y13139, and with nucleotide ID number 1302481.
 - SEQ ID NO. 2: The amino acid sequence of the suggested open reading frame YNR008w from Saccharomyces cerevisiae.
- SEQ ID NO. 3: Genomic DNA sequence of the Schizosaccharomyces pombe gene SPBC776.14.
 - SEQ ID NO. 4: Genomic DNA sequence of part of the Arabidopsis thaliana locus with GenBank accession number AB006704.
 - SEQ ID NO. 5: Nucleotide sequence of the Arabidopsis thaliana cDNA clone with GenBank accession number T04806, and nucleotide ID number 315966.
- SEQ ID NO. 6: Predicted amino acid sequence of the Arabidopsis thaliana cDNA clone with GenBank accession number T04806.
 - SEQ ID NO. 7: Nucleotide and amino acid sequence of the Zea mays EST clone with GenBank accession number Al491339, and nucleotide ID number 4388167.
- 25 SEQ ID NO. 8: Predicted amino acid sequence of the Zea mays EST clone with GenBank accession number AI491339, and nucleotide ID number 4388167.
- SEQ ID NO. 9: DNA sequence of part of the Neurospora crassa EST clone W07G1, with GenBank accession number Al398644, and nucleotide ID number 4241729.
 - SEQ ID NO. 10: Genomic DNA sequence of part of the Arabidopsis thaliana locus

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with GenBank accession number AC004557.

SEQ ID NO. 11: Genomic DNA sequence of part of the Arabidopsis thaliana locus with GenBank accession number AC003027.

SEQ ID NO. 12: DNA sequence of part of the Lycopersicon esculentum cDNA clone with GenBank accession number Al486635.

SEQ ID NO. 13: Amino acid sequence of the Schizosaccharomyces pombe putative open reading frame CAA22887 of the Schizosaccharomyces pombe gene SPBC776.14.

SEQ ID NO. 14: Amino acid sequence of the Arabidopsis thaliana putative open reading frame AAC80628 derived from the Arabidopsis thaliana locus with GenBank accession number AC004557.

SEQ ID NO 15: Amino acid sequence of the Arabidopsis thaliana putative open reading frame AAD10668 derived from the Arabidopsis thaliana locus with GenBank accession number AC003027.

Further provisional and/or partial sequences are defined through the following SEQ IDs:

SEQ ID NO. 16: The amino acid sequence of the yeast ORF YNR008w from Saccharomyces cerevisiae.

SEQ ID NO. 17: Amino acid sequence of the region of the Arabidopsis thaliana genomic sequence (AC004557).

SEQ ID NO. 18: Amino acid sequence of the region of the *Arabidopsis thaliana* genomic sequence (AB006704).

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SEQ ID NO. 19: The corresponding genomic DNA sequence and amino acid sequence of the yeast ORF YNROO8w from Saccharomyces cerevisiae.

SEQ ID NO. 20: The amino acid sequence of the yeast ORF YNROO8w from Saccharomyces cerevisiae derived form the corresponding genomic DNA sequence.

SEQ ID NO. 21: Genomic DNA sequence of the Saccharomyces cerevisiae
10 PDAT gene, YNR008w, genebank nucleotide ID number 1302481, and the suggested YNR008w amino acid sequence.

SEQ ID NO. 22: The suggested amino acid sequence of the yeast gene YNR008w from Saccharomyces cerevisiae.

SEQ ID NO. 23: Genomic DNA sequence of the Schizosaccharomyces pombe gene SPBC776.14.

SEQ ID NO. 24: Genomic DNA sequence of part of the Arabidopsis thaliana locus with genebank accession number AB006704.

SEQ ID NO. 25: Nucleotide sequence and the corresponding amino acid sequence of the *Arabidopsis thaliana* EST-clone with genebank accession number T04806, and ID number 315966.

SEQ ID NO. 26: Nucleotide and amino acid sequence of the Zea mays cDNA clone with genebank ID number 4388167.

SEQ ID NO. 27: Amino acid sequence of the Zea mays cDNA clone with genebank ID number 4388167.

SEQ ID NO. 28: DNA sequence of part of the Neurospora crassa cDNA clone WO7G1, ID number 4241729.

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SEQ ID NO. 29: Genomic DNA sequence of part of the Arabidopsis thaliana locus with genebank accession number AC004557.

5 SEQ ID NO. 30: Genomic DNA sequence of part of the Arabidopsis thaliana locus with genebank accession number AC003027.

SEQ ID NO. 31: DNA sequence of part of the Lycopersicon esculentum cDNA clone with genebank accession number Al486635.

<u>Claims</u>

- 1. An enzyme catalysing in an acyl-CoA-independent reaction the transfer of fatty acids from phospholipids to diacylglycerol in the biosynthetic pathway for the production of triacylglycerol.
- 2. An enzyme according to claim 1, comprising an amino acid sequence as set forth in SEQ ID No. 2 or a functional fragment, derivate, allele, homolog or isoenzyme thereof.

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- 3. An enzyme according to claims 1 or 2 designated as phospholipid:diacylglycerol acyltransferase (PDAT).
- 4. An enzyme according to claims 1 to 3, comprising an amino acid sequence as set forth in SEQ ID No. 16, 20 or 22 or a functional fragment, derivate, allele, homolog or isoenzyme thereof.
- 5. An enzyme according to claims 1 to 4, comprising an amino acid sequence selected from the group consisting of sequences as set forth in SEQ ID No. 6, 8, 13, 14, 15, 17, 18, 25 or 27 or a functional fragment, derivate, allele, homolog or isoenzyme thereof.
- 6. An enzyme according to claims 1 to 5, comprising an amino acid sequence encoded through a nucleotide sequence, a portion, derivate, allele or homolog thereof selected from the group consisting of sequences as set forth in SEQ ID No. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 or 31 or a functional fragment, derivate, allele, homolog or isoenzyme of the enzyme encoding amino acid sequence.
- 7. A nucleotide sequence encoding an enzyme catalysing in an acyl-CoAindependent reaction the transfer of fatty acids from phospholipids to diacylglycerol in the biosynthetic pathway for the production of triacylglycerol.

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- 8. A nucleotide sequence according to claim 7 encoding an enzyme designated as phospholipid:diacylglycerol acyltransferase (PDAT).
- 9. A nucleotide sequence according to claims 7 or 8, selected from the group consisting of sequences as set forth in SEQ ID No. 1, 3, 4, 10, 11, 19, 21, 23, 24, 29 or 30 or a portion, derivate, allele or homolog thereof.
- 10. A partial nucleotide sequence corresponding to a fullength nucleotide sequence according to claims 7 to 9, selected from the group consisting of sequences as set forth in SEQ ID No. 5, 7, 9, 12, 25, 26, 28 or 31 or a portion, derivate, allele or homolog thereof.
 - 11. A nucleotide sequence according to claims 7 to 10, comprising a nucleotide sequence which is at least 40% homologous to a nucleotide sequence selected form the group consisting of those sequences set forth in SEQ ID No. 1, 3, 4, 5, 7, 9, 10, 11, 12, 19, 21, 23, 24, 25, 26, 28, 29, 30 or 31.
- 12. A gene construct comprising a nucleotide sequence according to claims 7 to 11 operably linked to a heterologous nucleic acid.
 - 13. A vector comprising a nucleotide sequence according to claims 7 to 11 or a gene construct according to claim 12.
- 25 14. A vector according to claim 13, which is an expression vector.
 - 15. A vector according to claims 13 or 14, further comprising a selectable marker gene and/or nucleotide sequences for the replication in a host cell or the integration into the genome of the host cell.

16. A transgenic cell or organism containing a nucleotide sequence according to claims 7 to 11 and/or a gene construct according to claim 12 and/or a vector according to claims 13 to 15.

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- 17. A transgenic cell or organism according to claim 16 which is an eucaryotic cell or organism.
- 18. A transgenic cell or organism according to claims 16 or 17 which is a yeast cell or a plant cell or a plant.
 - 19. A transgenic cell or organism according to claims 16 to 18 having an altered biosynthetic pathway for the production of triacylglycerol.
 - 20. A transgenic cell or organism according to claims 16 to 19 having an altered oil content.
- 21. A transgenic cell or organism according to claims 16 to 20 wherein the activity of PDAT is altered.
 - 22. A transgenic cell or organism according to claims 16 to 21 wherein the altered activity of PDAT is characterized by an alteration in gene expression, catalytic activity and/or regulation of activity of the enzyme.
 - 23. A transgenic cell or organism according to claims 16 to 22 wherein the altered biosynthetic pathway for the production of triacylglycerol is characterized by the prevention of accumulation of undesirable fatty acids in the membrane lipids.
 - 24. A process for the production of triacylglycerol, comprising growing a transgenic cell or organism according to claims 16 to 23 under conditions whereby the said nucleotide sequence according to claims 7 to 11 is expressed.
 - 25. Triacylglycerols produced by a process according to claim 24.

- 26. Use of a nucleotide sequence according to claims 7 to 11 and/or an enzyme according to claims 1 to 6 for the production of triacylglycerol and/or triacylglycerols with uncommon fatty acids.
- 27. Use of a nucleotide sequence according to claims 7 to 11 and/or an enzyme according to claims 1 to 6 for the transformation of any cell or organism in order to be expressed in this cell or organism and result in an altered, preferably increased oil content of this cell or organism.

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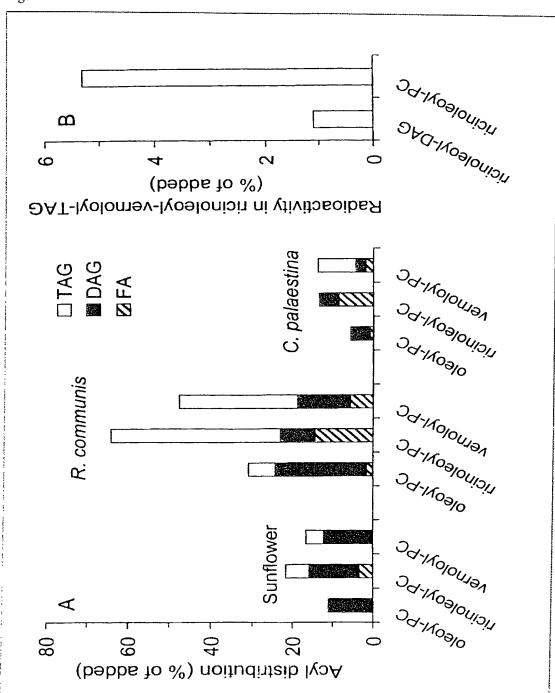
Abstract of the Disclosure

The present invention relates to the isolation, identification and characterization of nucleotide sequences encoding an enzyme catalysing the transfer of fatty acids from phospholipids to diacylglycerol in the biosynthetic pathway for the production of triacylglycerol, to the said enzymes and a process for the production of triacylglycerols.

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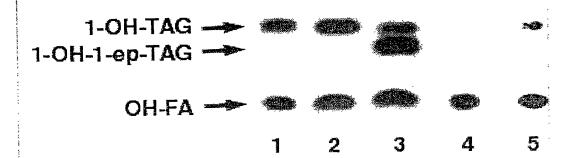
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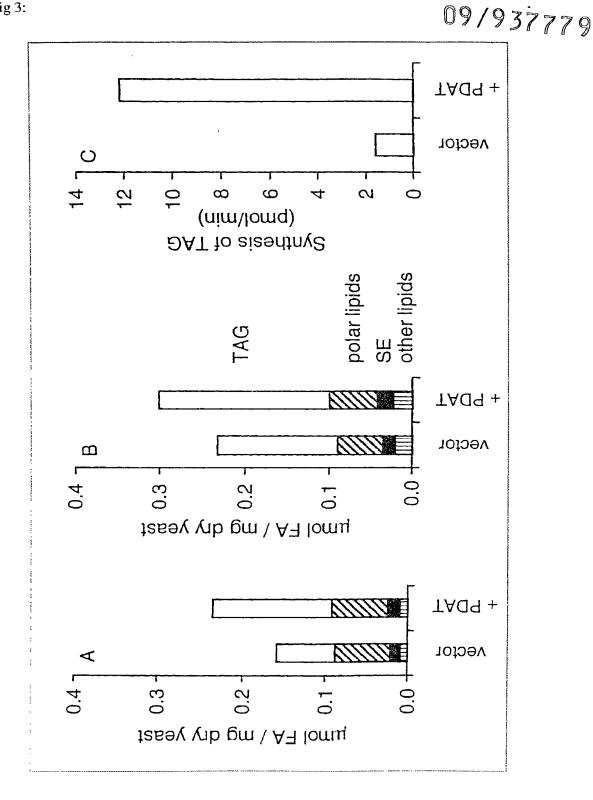
Fig. 1:



12.09.2000 **09/937779**





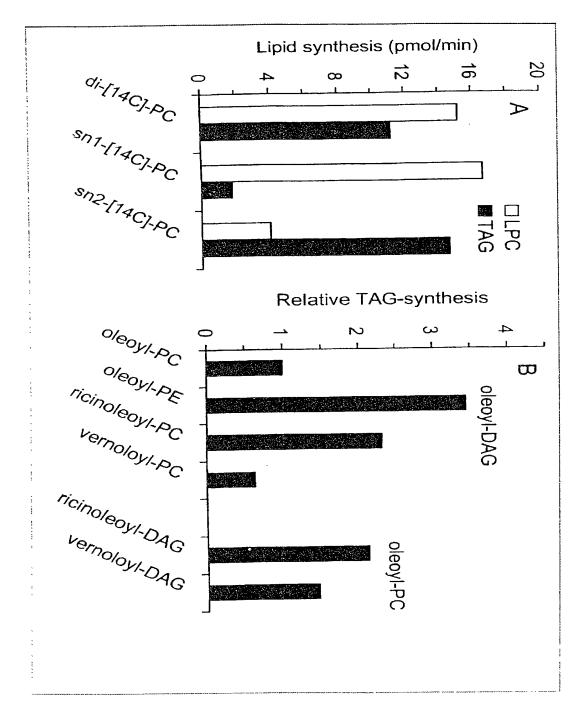


BASF-NAE 3377 / 99 PCT Fig 3:

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Fig.4:

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BASF-NAE 3377 / 99 PCT

Tables Tab. 1:

			mol % of adde	mol % of added [14C] -acyl group in TAG(1)	o in TAG		w
Substrate added [14C]-lipid ⁽²⁾ unlabelled lipid ⁽²⁾	unlabelled lipid ⁽²⁾	1-OH-TAG	2-OH-TAG	1-OH-1-ver-TAG 1-OH-2-ver-TAG	1-OH-2-ver-TAG	3-OH-TAG	
A mono-l' ¹⁴ C]-ricinoleoyl-DAG mono-ricinoleoyl-DAG	mono-ricinoleoyl-DAG	2,8	12,4				
A mono-[14C]-ricinoleoyl-DAG mono-vernoleoyl-DAG	mono-vernoleoyl-DAG	3,2	12,1	1,3	-	1	
A mono-[14C]-ricinoleoyl-DAG di-vernoleoyl-DAG	di-vernoleoyl-DAG	4	10	0,5	1,2		
A mono-[14C]-ricinoleoyl-DAG di-ricinoleoyl-PC	di-ricinoleoyl-PC	· c'0	24,8				
B mono-[¹⁴ C]-ricinoleoyl-PC	none	6,8	0,8		; ;	4,7	
C mono-[¹⁴ C]-ricinoleoyl-PC	di-oleoyl-DAG	8,6	8°, 0°			5,0	
C mono-[¹⁴ C]-ricinoleoyl-PC	mono-ricinoleoyl-DAG	5,7	16,7	t	1		
C mono-[14C]-ricinoleoyl-PC	di-ricinoleoyi-DAG	4,5	9,4	i i	ı	9,5	
C mcno-[¹⁴ C]-ricinoleoyl-PC	mono-vernoleoyl-DAG	0'9	11,5	10,9	0,5	7,4	
G mono-[14G]-ricinoleoyl-PC	di-vernoleoyl-DAG	2,9	10,8		8,4	6,8	
	-						

Tab. 2:

T1 plant	T2 plant number	nmol fatty acids per mg see	ed standard deviation
32-4	1	1277	±11 (n=2)
	4	1261	±63 (n=3)
	5	1369	+17 (n=3)
	6	1312	±53 (n=4)
	7	1197	±54 (n=5)
	8	1240	+78 (n=4)
	9	1283	$\pm 54 \text{ (n= 5)}$
	10	1381	$\pm 35 \text{ (n=5)}$
26-14	1	1444	±110 (n=4)
	2	1617*	+109 (n=4)
	3	1374	±37 (n=2)
	5	1562*	\pm 70 (n=4)
	6	1393	\pm 77 (n=4)
	7	1433	±98 (n=4)
	8	1581*	±82 (n=4)

Declaration, Power of Attorney and Petition

00474

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JUL 0 2 2002

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We (I), the undersigned inventor(s), hereby declare(s) that

My residence, post office address and citizenship are as stated below next to my name,

We (I) believe that we are (I am) the original, first, and joint (sole) inventor(s) of the subject matter which is claimed and for which a patent is sought on the invention entitled

NEW CLASS OF ENZYMES IN THE BIOSYNTHETIC PATHWAY FOR THE PRODUCTION OF TRIACYCLOGLYCEROL AND RECOMBINANT DNA MOLECULES ENCODING THESE ENZYMES

the specification of which

[]	is attached hereto.	
[x]	was filed onSeptember 28, 2001	as
	Application Serial No. <u>09/937,779</u>	
	and amended on	•
[x]	was filed as PCT international application	
	Number <u>PCT/EP/00/02701</u>	_
	on <u>March 28, 2000</u>	_
	and was amended under PCT Article 19	
	on (if applicable	le)

We (I) hereby state that we (I) have reviewed and understand the contents of the above—identified specification, including the claims, as amended by any amendment referred to above.

We (I) acknowledge the duty to disclose information known to be material to the patentability of this application as defined in Section 1.56 of Title 37 Code of Federal Regulations.

We (I) hereby claim foreign priority benefits under 35 U.S.C. § 119(a)—(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed. Prior Foreign Application(s)

Application No.	Country	Day/Month/Year	Priority Claimed
99106656.4	Europe	01 April 1999	[x] Yes [] No
99111321.8	Europe	10 June 1999	[x] Yes [] No
60/180687	United States of America	07 February 2000	[x] Yes [] No

Page 3 of 4 Declaration

0093/000003

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Page 2 of 4 Declaration

0093/000003

(Application	Number)	(Filing Date)
(Application	Number)	(Filing Date)
nternational application designat of this application is not disclosed irst paragraph of 35 U.S.C. § 112,	ing the United States, listed below a in the prior United States or PCT In I acknowledge the duty to disclose in	United States application(s), or § 365(c) of any PC and, insofar as the subject matter of each of the claim atternational application in the manner provided by the formation which is material to patentability as defined prior application and the national or PCT Internation
Application Serial No.	Filing Date	Status (pending, patented, abandoned)
		on Number 18,967; and RUSSEL E. WEINKAU

false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing

We (I) hereby claim the benefit under Title 35, United States Codes, § 119(e) of any United States provisional

thereon.

Page 4 of 4 Declaration

0093/000003

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.

SEQUENCE LISTING

<110> Dahlquist, Anders, Stahl, Ulf Lenman, Marit Banas, Antoni Ronne, Hans Stymne, Sten

<120> A NEW CLASS OF ENZYMES IN THE BIOSYNTHETIC PATHWAY FOR THE PRODUCTION OF
TRIACYLGLYCEROL AND RECOMBINANT DNA MOLECULES ENCODING THESE ENZYMES

<130> BASF-NAE-3377-99-Sept-2000

<140> US 09/937,779

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<151> 2000-03-23

<160> 31

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- Ser Glu Gln Pro Ser Ala Ser Glu Thr Gln Ser Val Ser Asn Lys Ser
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- Arg Lys Ser Lys Phe Gly Lys Arg Leu Asn Phe Ile Leu Gly Ala Ile 50 55 60
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- Val Phe Asp Pro Ala Thr Leu Asp Lys Phe Gly Asn Met Leu Gly Ser 85 90 95
- Ser Asp Leu Phe Asp Asp Ile Lys Gly Tyr Leu Ser Tyr Asn Val Phe 100 105 110
- Lys Asp Ala Pro Phe Thr Thr Asp Lys Pro Ser Gln Ser Pro Ser Gly 115 120
- Asn Glu Val Gln Val Gly Leu Asp Met Tyr Asn Glu Gly Tyr Arg Ser 130 135 140
- Ser Trp Ser Phe Asn Asn Cys Ser Ile Pro Tyr Phe Arg Lys Arg Leu 165 170 175
- Trp Gly Ser Trp Ser Met Leu Lys Ala Met Phe Leu Asp Lys Gln Cys 180 185 190
- Trp Leu Glu His Leu Met Leu Asp Lys Lys Thr Gly Leu Asp Pro Lys 195 200 205
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- Ile Gly Tyr Glu Pro Asn Asn Met Leu Ser Ala Ser Tyr Asp Trp Arg 245 250 255
- Leu Ser Tyr Ala Asn Leu Glu Glu Arg Asp Lys Tyr Phe Ser Lys Leu 260 265 270
- Lys Met Phe Ile Glu Tyr Ser Asn Ile Val His Lys Lys Lys Val Val

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Tyr Pro Ile His Lys Lys Ser Gly Gly Trp Phe Arg Leu Trp Phe Asp 65 70 75 80

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Met Leu Tyr Tyr Asp Pro Asp Leu Asp Asp Tyr Gln Asn Ala Pro Gly
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Val Gln Thr Arg Val Pro His Phe Gly Ser Thr Lys Ser Leu Leu Tyr
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Leu Asp Pro Arg Leu Arg Asp Ala Thr Ser Tyr Met Glu His Leu Val 130 135 140

Lys Ala Leu Glu Lys Lys Cys Gly Tyr Val Asn Asp Gln Thr Ile Leu 145 150 155 160

Gly Ala Pro Tyr Asp Phe Arg Tyr Gly Leu Ala Ala Ser Gly His Pro 165 170 175

Ser Arg Val Ala Ser Gln Phe Leu Gln Asp Leu Lys Gln Leu Val Glu 180 185 190

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- Trp Gly Gly Thr Ile Ser Gln Met Lys Thr Phe Ala Ser Gly Asn Thr 245 250 255
- Leu Gly Val Pro Leu Val Asn Pro Leu Leu Val Arg Arg His Gln Arg 260 265 270
- Thr Ser Glu Ser Asn Gln Trp Leu Leu Pro Ser Thr Lys Val Phe His 275 280 285
- Asp Arg Thr Lys Pro Leu Val Val Thr Pro Gln Val Asn Tyr Thr Ala 290 295 300
- Tyr Glu Met Asp Arg Phe Phe Ala Asp Ile Gly Phe Ser Gln Gly Val 305 310 315 320
- Val Pro Tyr Lys Thr Arg Val Leu Pro Leu Thr Glu Glu Leu Met Thr 325 330 335
- Pro Gly Val Pro Val Thr Cys Ile Tyr Gly Arg Gly Val Asp Thr Pro 340 345 350
- Glu Val Leu Met Tyr Gly Lys Gly Gly Phe Asp Lys Gln Pro Glu Ile 355 360 365
- Leu Lys Val Asp Ser Leu Asn Thr Val Glu Ile Asp Gly Val Ser His 385 390 395 400
- Thr Ser Ile Leu Lys Asp Glu Ile Ala Leu Lys Glu Ile Met Lys Glu 405 410 415
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 35 40 45
- Phe Ala Ser Thr Gln Leu Arg Ala Trp Ser Ile Leu Asp Cys Pro Tyr 50 55 60
- Thr Pro Leu Asp Phe Asn Pro Leu Asp Leu Val Trp Leu Asp Thr Thr 65 70 75 80

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Gly	Leu	Ser 115	Ala	Ile	Thr	Glu	Leu 120	Asp	Pro	Gly	Tyr	Ile 125	Thr	Gly	Pro
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Ala	His 370	Arg	Ser	Ser	Leu	Ala 375	Gly	Phe	Leu	Leu	Tyr 380	His	Asp	Asp	Pro
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Ile Tyr Glu Thr Glu Gly Ser Leu Val Ser Arg Ser Gly Thr Val Val 435 440 445

Asp Gly Asn Ala Gly Pro Ile Thr Gly Asp Glu Thr Val Pro Tyr His 450 455 460

Ser Leu Ser Trp Cys Lys Asn Trp Leu Gly Pro Lys Val Asn Ile Thr 465 470 475 480

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Lys Arg Asp Gly Asn Gly Arg Lys Arg Trp Arg Asp Ser Arg Arg Leu 65 70 75 80

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Gly Ala Tyr His Val His Asn Ser Asp Ser Asp Leu Phe Asp Asn Phe 100 105 110

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Ala 625	Glu	Leu	Asn	Asp	Tyr 630	Ile	Leu	Lys	Ile	Ala 635	Ser	Gly	Asn	Gly	Asp 640
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Thr	Lys	Ser	Leu 100	Leu	Tyr	Leu	Asp	Pro 105	Arg	Leu	Arg	Asp	Ala 110	Thr	Ser
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Thr 305	Glu	Glu	Leu	Met	Thr 310	Pro	Gly	Val	Pro	Val 315	Thr	Cys	Ile	Tyr	Gly 320
Arg	Gly	Val	Asp	Thr 325	Pro	Glu	Val	Leu	Met 330	Tyr	Gly	Lys	Gly	Gly 335	Phe

Asp Lys Gln Pro Glu Ile Lys Tyr Gly Asp Gly Asp Gly Thr Val Asn 340 345 350

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Asp Gly Leu Phe Arg Lys Arg Leu Trp Gly Gly Thr Phe Leu Cys Trp 35 40 45

Val Glu His Met Ser Leu Asp Asn Glu Thr Gly Leu Asp Pro Ala Gly 50 55 60

Ile Arg Val Arg Ala Val Ser Gly Leu Val Ala Ala Asp Tyr Phe Ala 65 70 75 80

Pro Gly Tyr Phe Val Trp Ala Val Leu Ile Ala Asn Leu Ala His Ile 85 90 95

Gly Tyr Glu Glu Lys Asn Met Tyr Met Ala Ala Tyr Asp Trp Arg Leu 100 105 110

Ser Phe Gln Asn Thr Glu Arg Asp Gln Thr Leu Ser Arg Met Lys Ser 115 120 125

Asn Ile Glu Leu Met Val Ser Thr Asn Gly Gly Lys Lys Ala Val Ile 130 135 140

Val Pro His Ser Met Gly Val Leu Tyr Phe Leu His Phe Met Lys Trp 145 150 155 160

Val Glu Ala Pro Ala Pro Leu Gly Gly Gly Gly Pro Asp Trp Cys 165 170 175

Ala Lys Tyr Ile Lys Ala Val Met Asn Ile Gly Gly Pro Phe Leu Gly 180 185 190

Val Pro Lys Ala Val Ala Gly Leu Phe Ser Ala Glu Ala Lys Asp Met 195 200 205

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			cgt ggc aaa Arg Gly Lys 60		
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Met Val Pro			att gaa agc Ile Glu Ser		
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		gaa Glu 275														864
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		gag Glu														1248
		cca Pro														1296
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cga Arg	ttt Phe 450	gaa Glu	agg Arg	aat Asn	acg Thr	agc Ser 455	gat Asp	gct Ala	ttc Phe	aac Asn	aaa Lys 460	aat Asn	ttg Leu	aca Thr	atg Met	1392
aaa Lys 465	gac Asp	gcc Ala	att Ile	aac Asn	atg Met 470	aca Thr	tta Leu	tcg Ser	ata Ile	tca Ser 475	cct Pro	gaa Glu	tgg Trp	ctc Leu	caa Gln 480	1440
		gta Val														1488

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aac gaa cac ata g Asn Glu His Ile A 355		sn Ala Ala Gly		1104
gct cca aag gca g Ala Pro Lys Ala V 370				1152
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tca gag gat gca t Ser Glu Asp Ala L 435		hr Asp Thr Tyr		1344
cga ttt gaa agg a Arg Phe Glu Arg A 450				1392
aaa gac gcc att a Lys Asp Ala Ile A 465				1440
aga aga gta cat g Arg Arg Val His G 4				1488
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	tcc Ser															1680
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	tgt Cys															1776
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Tyr 145	Ser	Thr	Ser	Ser	Leu 150	Asp	Asp	Leu	Ser	Glu 155	Asn	Phe	Ala	Val	Gl _y 160
Lys	Gln	Leu	Leu	Arg 165	Asp	Tyr	Asn	Ile	Glu 170	Ala	Lys	His	Pro	Val 175	Va]
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Ile Gln Leu Asn Thr Leu Ala Met Tyr Gly Leu Glu Lys Phe Phe Ser Arg Ile Glu Arg Val Lys Met Leu Gln Thr Trp Gly Gly Ile Pro Ser 405 410 Met Leu Pro Lys Gly Glu Glu Val Ile Trp Gly Asp Met Lys Ser Ser Ser Glu Asp Ala Leu Asn Asn Asn Thr Asp Thr Tyr Gly Asn Phe Ile 440 Arg Phe Glu Arg Asn Thr Ser Asp Ala Phe Asn Lys Asn Leu Thr Met 455 Lys Asp Ala Ile Asn Met Thr Leu Ser Ile Ser Pro Glu Trp Leu Gln 470 475 Arg Arg Val His Glu Gln Tyr Ser Phe Gly Tyr Ser Lys Asn Glu Glu Glu Leu Arg Lys Asn Glu Leu His His Lys His Trp Ser Asn Pro Met 500 Glu Val Pro Leu Pro Glu Ala Pro His Met Lys Ile Tyr Cys Ile Tyr 520 Gly Val Asn Asn Pro Thr Glu Arg Ala Tyr Val Tyr Lys Glu Glu Asp 535 Asp Ser Ser Ala Leu Asn Leu Thr Ile Asp Tyr Glu Ser Lys Gln Pro Val Phe Leu Thr Glu Gly Asp Gly Thr Val Pro Leu Val Ala His Ser 570 Met Cys His Lys Trp Ala Gln Gly Ala Ser Pro Tyr Asn Pro Ala Gly Ile Asn Val Thr Ile Val Glu Met Lys His Gln Pro Asp Arg Phe Asp 600 Ile Arg Gly Gly Ala Lys Ser Ala Glu His Val Asp Ile Leu Gly Ser Ala Glu Leu Asn Asp Tyr Ile Leu Lys Ile Ala Ser Gly Asn Gly Asp 630 Leu Val Glu Pro Arg Gln Leu Ser Asn Leu Ser Gln Trp Val Ser Gln 650 Met Pro Phe Pro Met 660

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						cat His										960
						tct Ser										1008
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- Lys Arg Asp Gly Asn Gly Arg Lys Arg Trp Arg Asp Ser Arg Arg Leu 65 70 75 80
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- Gly Ala Tyr His Val His Asn Ser Asp Ser Asp Leu Phe Asp Asn Phe 100 105 110
- Val Asn Phe Asp Ser Leu Lys Val Tyr Leu Asp Asp Trp Lys Asp Val 115 120 125
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- Tyr Ser Thr Ser Ser Leu Asp Asp Leu Ser Glu Asn Phe Ala Val Gly 145 150 155 160
- Lys Gln Leu Leu Arg Asp Tyr Asn Ile Glu Ala Lys His Pro Val Val
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- Met Val Pro Gly Val Ile Ser Thr Gly Ile Glu Ser Trp Gly Val Ile 180 185 190
- Gly Asp Asp Glu Cys Asp Ser Ser Ala His Phe Arg Lys Arg Leu Trp 195 200 205
- Gly Ser Phe Tyr Met Leu Arg Thr Met Val Met Asp Lys Val Cys Trp 210 215 220
- Leu Lys His Val Met Leu Asp Pro Glu Thr Gly Leu Asp Pro Pro Asn 225 230 235 240
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Ile Asn Val Thr Ile Val Glu Met Lys His Gln Pro Asp Arg Phe Asp 595 600 605

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Ala Glu Leu Asn Asp Tyr Ile Leu Lys Ile Ala Ser Gly Asn Gly Asp 625 630 635 640

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Pro Ser Glu Glu Val Val His Asp Glu Asp Ser Gln Lys Lys Pro His
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Glu Ser Ser Lys Ser His His Lys Xaa Ser Asn Gly Gly Lys Trp
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Ser Cys Ile Asp Ser Cys Cys Trp Phe Ile Gly Cys Val Cys Val Thr
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Trp Trp Phe Leu Leu Phe Leu Tyr Asn Ala Met Pro Ala Ser Phe Pro
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09/937779 JC05 Rec'd PCT/PTO 2 8 SEP 2007

SEQUENCE LISTENING

<110> BASF AG

<120> A NEW CLASS OF ENZYMES IN THE BIOSYNTHETIC PATHWAY FOR THE PRODUCTION OF TRIACYLGLYCEROL AND RECOMBINANT DNA MCLECULES ENCODING THESE ENZYMES

<130> EASF-NAE-3377-99-Sept-2000

<140> PCT/EP 00/02701

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<170> PatentIn Ver. 2.1

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Lys Arg Asp Gly Asn Gly Arg Lys Arg Trp Arg Asp Ser Arg Arg Leu
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	tat Tyr														864
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10/58

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Pro	Val	Leu	Ser 580	Ala	Gly	Tyr	Met	Cys 585	Ala	Lys	Ala	qrT	Arg 590	Gly	ĽУъ
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12.09.2000

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Thr	Ser	Ile	Leu	Lys 405	Asp	Glu	Ile	Ala	Leu 410	Lys	Glu	Ile	Met	Lys 415	Gln
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His	Phe 290	Ser	Gly	Gly	Ala	Ala 295	Lys	Lys	ązA	Lys	Arg 300	Va1	Tyr	His	Cys
Asp 305	Glu	Glu	Glu	Tyr	Gln 310	Ser	Lys	Tyr	Ser	Gly 315	Trp	Pro	Thr	Asn	īle 320
Ile	Asn	Ile	Glu	Ile 325	Pro	Ser	Thr	Ser	Ala 330	Arg	Glu	Leu	Ala	Asp 335	Gly
Thr	Leu	Phe	Lys 340	Ala	Ile	Glu	Asp	Tyr 345	Asp	Pro	Asp	Ser	Lys 350	Arg	Met
Leu	His	Gln 355	Leu	Lys	Lys	Tyr	Val 360	Pro	Phe	Phe	Va1	Ile 365	Arg	Asn	Ile
Ala	His 370	Arg	Ser	Ser	Leu	Ala 375	Gly	Phe	Leu	Leu	Tyr 380	Hıs	Asp	Asp	Pro
Val 385		Asn	Pro	Leu	Thr 390	Pro	Trp	Glu	Arg	Pro 395	Pro	Ile	Lys	Asn	Val 400
Phe	Cys	Ile	Tyr	Gly 405	Ala	His	Leu	Lys	Thr 410	Glu	Val	Gly	ŢŊï	Tyr 415	Phe
Ala	Pro	Ser	Gly 420	Lys	Fro	Tyr	Pro	Asp 425	Asn	Trp	Ile	Ile	Thr 430	qaA	Ile
Ile	Tyr	Glu 435	Thr	Clu	GīA	Ser	Leu 440	Val	Ser	Яrg	Ser	Gly 445	Thr	Vai	Val
Asp	Gly 450	Asn	Ala	Gly	Pro	Ile 455	Thr	Gly	qaA	Glu	Thr 460	Val	Pro	Tyr	His
Ser 465		Ser	Trp	Cys	Lys 470	Asn	Trp	Leu	Gly	Pro 475	Lys	Val	Asn	Ile	Thr 480
Met	Ala	Pro	Gln	Ile 485	Leu	Ile	Gly	Lys	Ile 490	Lys	Gln	Gln	Pro	Glu 495	His
Asp	Gly	Ser	Asp 500	Val	His	Va1	Glu	Leu 505	Asn	Val	Asp	His	Glu 510	His	Gly
Ser	Asp	Ile 515	Ile	Ala	Asn	Мег	Thr 520	Lys	Ala	Pro	Arg	Val 525	Lys	Tyr	Ile
Thr	Phe 530	Tyr	Glu	Asp	Ser	Glu 535	Ser	Ile	Pro	Gly	Lys 540	Arg	Thr	Ala	Val
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Ala	Tyr 290	Leu	Asp	Leu	Glu	Arg 295	Arg	Asp	Arg	Туг	Phe 300	Thr	Lys	Leu	Lys
Glu 305	Gln	Ile	Glu	Leu	Phe 310	His	Gln	Leu	Ser	Gly 315	Glu	Ĺys	Val	Cys	Leu 320
Ile	Gly	Hıs	Ser	Met 325	Gly	Ser	Gln	Ile	330 Ile	Phe	Tyr	Phe	Met	Lys 335	Trp
Val	Glu	Ala	Glu 340	G1y	Pro	Leu	Tyr	Gly 345	Asn	Gly	Gly	Arg	Gly 350	Trp	Va1
Asn	Glu	His 355	Ile	Asp	Ser	Phe	11e 360	Asn	Ala	Ala	Gly	Thr 365	Leu	Leu	Gly
Ala	Pro 370	Lys	Ala	Val	Pro	Ala 375	Leu	Ile	Ser	Gly	Glu 380	Met	Lys	Asp	Thr
Ile 385	Gln	Leu	Asn	Thr	Leu 390	Ala	Met	Tyr	G1y	Leu 395	Glu	Lys	Phe	Phe	Ser 400
Yrā	Ile	Glu	Arg	Val 405	Lys	Met	Leu	G1n	Thr 410	Trp	G1y	Gly	Ile	Pro 415	Ser
Мес	Leu	Pro	Lys 420	Gly	Glu	Glu	Val	Ile 425	Trp	Gly	Asp	Met	Lys 430	Ser	Ser
Ser	Glu	Asp 435	Ala	Leu	Asn	Asn	Asn 440	Thr	Asp	Thr	Tyr	Gly 445	Asn	Phe	Ile
Arg	Phe 450	Glu	Arg	Asn	Thr	Ser 455	Asp	Ala	Phe	Asn	Lys 460	Asn	Leu	Thr	Met
Lys 465	Asp	Ala	Ile	Asn	Мес 470	Thr	Leu	Ser	Ile	Ser 475	Pro	Glu	Trp	Leu	Gln 480
Ъrg	Arg	Val	Hıs	Glu 485	Gln	Tyr	Ser	Phe	G1y 490	ĵλι	Ser	Lys	Asn	Glu 495	Glu
Glu	Leu	Arg	Lys 500	Asn	Glu	Leu	His	His 505	rys	Hıs	Trp	Ser	Asn 510	Pro	Mec
Glu	Val	Pro 515	Leu	Pro	Glu	Ala	Pro 520	Hıs	Met	Ĺys	Ile	Туг 525	Cys	Ile	Tyr
Gly	Val 530	Asn	Asn	Pro	Thr	Glu 535	Arg	Ala	Tyr	Va1	Tyr 540	Lys	Glu	Glu	Asp
Asp 545	Ser	Ser	Ala	Leu	Asn 550	Leu	Thr	Ile	Asp	Tyr 555	Glu	Ser	Lys	Gln	Pro 560
Val	Phe	Leu	Thr	Glu 565	Gly	Asp	Gly	Thr	Val 570	Pro	Leu	Val	Ala	His 575	Ser
Met	Cys	Hıs	Lys 580	Trp	Ala	Gln	Gly	Ala 585	Ser	Pro	Tyr	Asn	Pro 590	Ala	Gly
Ile	Asn	Val 595	Thr	Ile	Val	Glu	Met 600	Lys	Hıs	Gln	Pro	Asp 605	Arg	Phe	qzA

Ile Arg Gly Gly Ala Lys Ser Ala Glu His Val Asp Ile Leu Gly Ser 610 615 620

Ala Glu Leu Asn Asp Tyr Ile Leu Lys Ile Ala Ser Gly Asn Gly Asp 625 630 635 640

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Met Pro Phe Pro Met 660

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Trp Cys Ser 35	Ser Trp	Leu Tyr	Pro I 40	ie His	Lys Lys	Ser 45	Gly	Gly	Trp					
Phe Arg Leu 50	Trp Phe	Asp Ala 55		al Leu	Leu Ser 60		Phe	Thr	Arg					
Cys Phe Ser 65	Asp Arg	Met Met 70	Leu T	Yr Tyr	Asp Pro	Asp	Leu	qzA	Asp 80					
Tyr Gln Asn	Ala Pro 85	Gly Val	Gln T	Thr Arg 90	Val Pro	His	Phe	Gly 95	Ser					
Thr Lys Ser	Leu Leu 100	Tyr Leu		ro Arg .05	Leu Arg	Asp	Ala 110	Thr	Ser					
Tyr Met Glu 115	Eis Leu	Val Lys	Ala L 120	eu Glu	Lys Lys	Cys 125	Gly	Tyr	Val					
Asn Asp Gln 130	Thr Ile	Leu Gly 135		Pro Tyr	Asp Phe	Arg	Tyr	Gly	Leu					
Ala Ala Ser 145	Gly His	Pro Ser 150	Arg V	al Ala	Ser Gln 155	Phe	Leu	Gln	Asp 160					
Leu Lys Gln	Leu Val 165	Glu Lys	Thr S	Ser Ser 170	Glu Asn	Glu	Gly	Lys 175	Pro					
Val Ile Leu	Leu Ser 180	His Ser		Gly Gly .85	Leu Phe	Val	Leu 190		Phe					
Leu Asn Arg 195	Thr Thr	Pro Ser	Trp A 200	rg Arg	Lys Tyr	Ile 205	Lys	His	Phe					
Val Ala Leu 210	Ala Ala	Pro Trp 215		ly Thr	Ile Ser 220		Met	Lys	Thr					
Phe Ala Ser 225	Gly Asn	Thr Leu 23J	Gly V	al Pro	Leu Val 235	Asn	Pro	Leu	Leu 240					
Val Arg Arg	His Gln 245	Arg Thr	Ser G	lu Ser 250	Asn Gln	Trp		Leu 255	Pro					
Ser Thr Lys	Val Phe 260	His Asp		hr Lys 65	Pro Leu		Val 270	Thr	Pro					
Gln Val Asn 275	Tyr Thr	Ala Tyr	Glu M 280	et Asp	Arg Phe	Phe 285	Ala	qaƙ	Ile					

Gly	Phe 290	Ser	Gln	Gly	Val	Val 295	Pro	Tyr	Lys	Thr	Arg 300	Val	Leu	Pro	Leu
Thr 305	Glu	Glu	Leu	Met	Thr 310	Pro	Gly	Val	Pro	Val 315	Thr	Cys	Ile	Tyr	Gly 320
Arg	Gly	Val	Asp	Thr 325	Pro	Glu	Val	Leu	Met 330	Tyr	Gly	Lys	Gly	Gly 335	Phe
Asp	Lys	Gln	Pro 340	Glu	Ile	Lys	Tyr	Gly 345	qsA	Gly	Asp	Gly	Thr 350	Val	Asn
Leu	Ala	Ser 355		Ala	Ala	Leu	Lys 360	Val	Asp	Ser	Leu	Asn 365	Thr	Va1	Glu
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Lys 385		Ile													

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Asp Gly		eu 35	Phe	Arg	Lys	Arg	Leu 40	Trp	Gly	Gly	Thr	Phe 45	Leu	Cys	Trp
Val Gl ⁻ 5		lis	Met	Ser	Leu	Asp 55	Asn	Glu	Thr	Gly	Leu 60	Asp	Pro	Ala	Gly
Ile Ar 65	ā ∧	/al	Arg	Ala	Val 70	Ser	Gly	Leu	Val	Ala 75	Ala	Asp	Ţγr	Phe	Ala 80
Pro Gl	y T	ľýľ	Phe	Val 85	Trp	Ala	Val	Leu	Ile 90	Ala	Asn	Leu	Ala	His 95	Ile
Gly Ty	r (Glu	Glu 100	Lys	Asn	Met	Tyr	Met 105	Ala	Ala	Tyr	Asp	Trp 110	Arg	Leu
Ser Pr		Gln 115	Asn	Thr	Glu	Arg	Asp 120	Gln	Thr	Leu	Ser	Arg 125	Met	Lys	Ser
Asn II	le 30	Glu	Lеч	Met	Val	Ser 135	Thr	Asr	ı Gly	Gly	- Lys 140	Lys	Ala	. Val	Ile
Val P: 145	ro	His	Ser	Met	Gly 150	Val	. Leu	ŢŢI	r Ph€	e Leu 155	His	. Phe	Met	Lys	Trp 160
Val G	lu	Ala	. Pro	Ala 165		Let	: Gly	Gly	7 Gly	y Gly	7 Gly	Pro	Asī	7rg	Cys
Ala L	УS	ŢŸŢ	11e	e Lys	ala	. Val	l Met	As:	n Ilo	e Gly	y Gly	Pro	Phe 190	e Lev	ı Gly
Val P	ro	Lys 195		a Val	Ala	Gly	/ Let 200	ı Ph	e Se	r Ala	a Glu	1 Ala 205	a Ly: 5	s As)	o Met
Arg M	let 10	The	r Arg	ולד ק	r Trr	21	p Sei 5	r Th	r M∈	t Se:	r Met 22(Lev	ı Pro	o Ly	s Gly
Gly A 225	qz	Thi	s Ile	e Tr	o G1 <u>y</u> 230	/ G1;	y Le:	ı As	p Tr	p Se 23	r Pro 5	o Gl	u Le	u Pr	o Asn 240
Ala E	Pro	Gl	ı Me	c Gli 24	u Ile 5	e Ty	r Se	r Le	u Ty 25	r Gl	y Val	l Gl	y Il	e Pr 25	o Thr 5
Glu A	\rg	Ala	a Ty: 26	r Va O	1 Ty:	r Ly	s Le	u As 26	n Gl 5	n Se	r Pr	o As	p S∈ 27	r Cy O	s Ile
Pro !	?he	G1:		e Ph	e Thi	r Se	r Al 28	a Hi O	s Gl	u Gl	u As	p Gl 28	u As 5	p Se	r Cys

Leu	Lvs	Ala	Gly	Val	Tyr	Asn	Val	Asp	Gly	qaA	Glu	Thr	Va1	Pro	Val
	290		-			295					300				

Leu Ser Ala Gly Tyr Met Cys Ala Lys Ala Trp Arg Gly Lys Thr Arg

Phe Asn Pro Ser Gly Tie Lys Thr Tyr Ile Arg Glu Tyr Asn His Ser 325 330 335

Pro Pro Ala Asn Leu Leu Glu Gly Arg Gly Thr Gln Ser Gly Ala His 340 345

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Gly Ile Phe Glu Trp 385

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								gtt Val 25								96
			Hıs					tta Leu						_		144
	-	-	_	-		_		gag Glu	_			_		-	-	192
		-			-			cgt Arg		_	_		_	_	_	240
					_			ggt Gly	_		-	_		_		288
								gat Asp 105								336
								tat Tyr								384
								att Ile								432
								ctc Leu								480
								atc Ile	_	-					-	528
								gga Gly 185								576

gga Gly	gac Asp	gat Asp 195	gag Glu	tgc Cys	gat Asp	agt Ser	tct Ser 200	gcg Ala	cat His	ttt Phe	cgt Arg	aaa Lys 205	cgg Arg	ctg Leu	tgg Trp	624
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ttg Leu 225	aaa Lys	cat Hıs	gta Val	atg Met	tta Leu 230	gat Asp	cct Pro	gaa Glu	aca Thr	ggt Gly 235	ctg Leu	gac Asp	cca Pro	ccg Pro	aac Asn 240	720
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ggc Gly	tat Tyr	gaa Glu 275	Pro	aat Asn	aaa Lys	atg Met	acg Thr 280	agt Ser	gct Ala	gcg Ala	tat Tyr	gat Asp 285	tgg Trp	agg Arg	ctt Leu	864
gca Ala	tat Tyr 290	Leu	gat Asp	cta Leu	gaa Glu	aga Arg 295	cgc Arg	gat Asp	agg Arg	tac Tyr	Ett Phe 300	Thr	aag Lys	cta Leu	aag Lys	912
gaa Glu 305	Gln	ato Ile	gaa Glu	. ctg Leu	ttt Phe 310	His	caa Gln	ttg Leu	agt Ser	ggt Gly 315	Giu	aaa Lys	gtt Val	tgt Cys	tta Leu 320	960
att Ile	gga Gly	cat His	tct Ser	atg Met 325	Gly	tct Ser	cag Gln	att Ile	ato le Ile	Ph∈	tac Tyr	ttt Phe	atg Met	aaa Lys 335	tgg Trp	1008
gto Val	gag I Glu	get LAla	gaa a Glu 340	ı Gly	cct Pro	ctt Leu	tac Tyr	ggt Gly 345	/ Asn	ggt Gly	ggt Gly	: cgt / Arg	350 350	Tr	gtt Val	1056
aac Asr	gaa n Glo	cac His	5 I16	e gat e Asp	t tca Ser	ttc Phe	att : Ile : 360	Asr	gca n Ala	gca Ala	: GJ7	g acg 7 Thr 365	Let	ctg Leu	ggc Gly	1104
gct Ala	cca Pro 370	Ly:	g gca s Ala	a gtt a Val	cca L Pro	gct Ala 375	. Leu	att i Ile	agt e Ser	ggt Gly	gaa Glu 380	ı Met	g aaa Lys	gat Asp	acc Thr	1152
att [16	e Glr	a tta n Lev	a aat ı Asr	acg n Thi	g tta Lev 390	ı Ala	ato Met	g tat Tyl	ggt Gly	: ttg / Let 395	ı GD	a aag 1 Lys	j tto s Phe	ttc Phe	tca Ser 400	1200
aga Arq	a att g īle	gaq e Gli	g aga u Arg	a gta Val 409	L Lys	atç Met	rtta Lei	e caa i Gl:	a acc n Thr 410	rr	o Gl;	t ggt / Gly	ata 7 Ile	e cca Pro 413	tca Ser	1248
aty Me	g cta t Lei	a cca ı Pro	aag o Lys 420	s Gly	a gaa / Glu	ı gaç ı Glu	g gto 1 Val	act 1 Tle 425	∋ Tr <u>:</u>	o Glž ā āāč	j ja: . Asi	acg Met	aag Lys 430	Ser	tct Ser	1296

		-	-						_					ttc Phe		1344
_					_			-					_	aca Thr	-	1392
	-							_				_		ctc Leu		1440
-		_												gaa Glu 495	_	1488
														cca Pro		1536
	_				-									ata Ile		1584
						-				-				gag Glu	_	1632
-			~	-		_			_		_	-	_	caa Gln		1680
_						_			-	_				cat His 575		1728
-														gcc Ala		1776
		_				_	_			_		-		ttc Phe		1824
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īle	Ser 50	Gly	Ser	Ala	Lys	Arg 55	Asn	Glu	Arg	Gly	Lys 60	Asp	Phe	Asp	Arg
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Ile	Phe	īle	Leu	Gly 85	Ala	Phe	Leu	Gly	Val 90	Leu	Leu	Pro	Phe	Ser 95	₽he
Gly	Ala	Туг	Hıs 100	Val	His	Asn	Ser	Asp 105	Ser	qaA	Leu	Phe	Asp 110	Asn	Phe
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Lys	Gln	. Leu	Leu	Arg 165		Tyr	Asn	ı Ile	e Glu 170	Ala	Lys	Hıs	Pro	Val 175	Val
Met	Va1	. Prc	Gly 180		īle	Ser	Thr	Gly 185	r Ile	e Glu	Ser	Trp	Gly 190	Val	Ile
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Leu 225		s His	s Val	. Mes	Leu 230	Asp	Pro	Glu	ı Thr	Gly 235	r Leu	Asp	Prc) Pro	240
Phe	e The	. Lev	ı Arç	Ala 245		Glr	ı Gly	/ Phe	e Glu 250	ı Ser	Thr	Asp	Tyr	Phe 259	e Ile
Ala	a Gly	/ Ty:	Tr; 260		e Trp	Asr	ı Lys	3 Val 265	i Phe	e Glr	n Asn	Leu	: Gly 270	v Val	Ile
Gly	/ Ty:	Glu 275) Asr	ı Lys	Мет	Th: 280	Sei	: Ala	i Alá	ı Tyr	Asp 285	Trp	Arg	j Leu

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Ile	Gly	His	Ser	Met 325	Gly	Ser	Gln	Ile	Ile 330	Phe	Tyr	Phe	Met	Lуs 335	Trp
Val	Glu	Ala	Glu 340	Gly	Pro	Leu	Tyr	Gly 345	Asn	Gly	Gly	Arg	Gly 350	Trp	Val
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Met	Leu	Pro	Lys 420	Gly	Glu	Glu	Va l	Ile 425	Trp	Gly	Asp	Mec	Lys 430	Ser	Ser
Ser	Glu	Asp 435	Ala	Leu	Asn	Asn	Asn 440	Thr	Asp	Thr	Tyr	Gly 445	Asn	Phe	Ile
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Lys 465	σεA	Ala	Ile	Asn	Met 470	Thr	Leu	Ser	Ile	Ser 475	Pro	Glu	Trp	Leu	Gln 480
Arg	Arg	Val	His	Glu 485	Gln	Tyr	Ser	Phe	Gly 490	Туг	Se≆	Lys	Asn	Glu 495	Glu
Glu	Leu	Arg	Lys 500	Asn	Glu	Leu	His	His 505	Lys	His	Trp	Ser	Asn 510	Pro	Met
Glu	Val	Pro 515	Leu	Pro	Glu	Ala	Prc 520	Hıs	Met	Lys	Ile	Tyr 525	Cys	īle	Tyr
Gly	Val 530	Asn	Asn	Pro	Thr	Glu 535	Arg	Ala	Tyr	Va1	Tyr 540	Lys	Glu	G1u	Asp
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Val	Fhe	Leu	Thr	Glu 565	Gly	Asp	Gly	Thr	Val 570	Pro	Leu	Val	Ala	Hıs 575	Ser
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50/58

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57/58

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